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(54) Title: **NEUROPEPTIDE Y RECEPTOR Y5 AND NUCLEIC ACID SEQUENCE**

(57) Abstract

The present invention provides novel NPY/PYY receptor proteins and the nucleic acid sequence encoding them. The invention is directed to the isolation, characterization, and pharmacological use of these receptors and nucleic acids. In particular, this invention provides human and rat NPY/PYY receptors (which we call the NPY Y5 receptor) and nucleic acids. Also provided are recombinant expression constructs useful for transfecting cells and expressing the protein *in vitro* and *in vivo*. The invention further provides methods for detecting expression levels of the protein as well as methods for screening for receptor antagonists and agonists to be used for the treatment of obesity or anorexia, respectively.

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Jonathan A. Bard, et al.
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NEUROPEPTIDE Y RECEPTOR Y5 AND NUCLEIC ACID SEQUENCE**BACKGROUND OF THE INVENTION****Field of the Invention**

This invention relates to a novel neurotransmitter Neuropeptide Y receptor, its nucleic acid sequence, and compounds, compositions, and methods for their use.

Summary of the Related Art

Neuropeptide Y (NPY) is a 36-amino acid peptide neurotransmitter that is located throughout the central and peripheral nervous systems. Tatemoto, *Proc. Natl. Acad. Sci. USA* 79, 5485 (1982); Hazlewood, *Proc. Soc. Exp. Biol. Med.* 202, 44 (1993). It affects a broad range of phenomena, including blood pressure regulation, memory, anxiolysis/sedation, food and water appetite, vascular and other smooth muscle activity, intestinal electrolyte secretion, and urinary sodium excretion. *E.g.*, Colmers and Wahlestedt, *The Biology of Neuropeptide Y and Related Peptides* (Humana Press, Totowa, NJ, 1993); Kalra et al., *Phys. & Behavior* 50, 5 (1991).

Peptide YY (PYY) is also a 36 amino acid peptide and has significant sequence homology (70%) to NPY. Tatemoto et al., *Nature* 296, 659 (1982). Its anatomical distribution is similar to that of NPY, although it is located mainly in the endocrine cells of the lower gastrointestinal tract. Bottcher et al., *Regul. Pept.* 8, 261 (1984). Like NPY, PYY stimulates feeding in rats. Morley et al., *Brain Res.* 341, 200 (1985). Along with the pancreatic polypeptide (PP), NPY and PYY have a common tertiary structure, characterized by the so-called PP-fold. Glover, *Eur. J. Biochem.* 142, 379 (1985). Both NPY and PYY show about a 50% sequence homology with PP.

Because of their structural similarities, NPY and PYY have a number of common receptors. At least four receptor subtypes, Y1, Y2, Y3, and Y4/PP, have been identified. The affinity for NPY, PYY, and various fragments thereof varies among the subtypes. *See, e.g.*, Bard et al. (WO 95/17906) and references cited therein. For example, Y1 and Y2 subtypes have high affinity for NPY and PYY. Whereas Y1 has high affinity for (Leu³¹Pro³⁴)NPY ((LP)NPY) and low affinity for (13-36)NPY, Y2 behaves oppositely. Y3 has high affinity for NPY but low affinity for PYY. Y4/PP has a high affinity for PP but relatively low affinity for NPY.

Wahlestedt (WO 93/24515) and Larhammar et al. (*J. Biol. Chem.* 267, 10935 (1992)) describe the cloning and identification of the human Y1-type NPY/PYY receptor isolated from

human fetal brain tissue. Selbie et al. (WO 93/09227) disclosed the full length cDNA sequence of the Y1 receptor from human hippocampus. Eva et al. (*FEBS Lett.* 271, 81 (1990)) cloned the NPY Y1 receptor from rat forebrain. Eva et al. (*FEBS Lett.* 314, 285 (1992)) cloned the NPY Y1 receptor from murine genomic DNA.

5 The Y2-type receptor has also been cloned. Gerald et al. (WO 95/21245) disclosed the cDNA sequence of human hippocampal Y2 and two rat Y2 clones. Rose et al. (*J. Biol. Chem.* 270, 22661 (1995)) disclosed the cDNA sequence of the Y2 receptor from a human neuroblastoma cell line.

Bard et al. (*supra*) and Lundell et al. (*J. Biol. Chem.* 270, 29123 (1995)) described
10 cloning the cDNA sequence of the Y4/PP receptor from both rat spleen and human placenta.

To date, the Y3 receptor has not been cloned.

Because of the important role of NPY and PYY in a number of physiological processes, such as feeding, there is a strong need to further develop materials and methods for investigating the mechanistic behavior of these compounds and for treating diseased and other
15 abnormal states associated with the physiological processes in which NPY and PYY act. Specifically, the NPY analogs/fragments that induce feeding, such as (LP)(3-36)NPY, do not bind to the previously identified NPY/PYY receptors with affinities consistent with the feeding response. Accordingly, there is a need and desire to identify the NPY/PYY receptor that is responsible for the feeding response. Antagonists to such a receptor could be used to treat
20 obesity and diabetes by reducing appetite and food consumption.

SUMMARY OF THE INVENTION

The present invention provides, *inter alia*, novel NPY/PYY receptor proteins. Also provided are the nucleic acid sequences encoding these novel receptor proteins, as well as compounds and methods for using these proteins and their nucleic acid sequences.

25 The present invention provides novel proteins, nucleic acids, and methods useful for developing and identifying compounds for the treatment of such diseases and disorders as obesity. Identified and disclosed herein is the protein sequence for a novel receptor for the neurotransmitters Neuropeptide Y (NPY) and Peptide YY (PYY) and the nucleic acid sequence encoding this receptor, which we call the NPY Y5 (or simply "Y5") receptor. The importance
30 of this discovery is manifested in the effects of NPY, which include blood pressure regulation, memory enhancement, anxiolysis/sedation, and increased food intake. Thus, this receptor

protein is useful for screening for NPY/PYY agonist and antagonist activity for controlling these conditions.

In one aspect of the present invention, we provide isolated nucleic acid sequences for a novel NPY and PYY receptor, the Y5 receptor. In particular, we provide the cDNA sequences encoding for the rat and human receptors and isoforms thereof. These nucleic acid sequences have a variety of uses. For example, they are useful for making vectors and for transforming cells, both of which are ultimately useful for production of the Y5 receptor protein. They are also useful as scientific research tools for developing nucleic acid probes for determining receptor expression levels, *e.g.*, to identify diseased or otherwise abnormal states. They are useful for developing analytical tools such as antisense oligonucleotides for selectively inhibiting expression of the receptor gene to determine physiological responses.

In another aspect of the present invention, we provide a homogenous composition comprising the receptor Y5 protein. The protein is useful for screening drugs for agonist and antagonist activity, and, therefore, for screening for drugs useful in regulating physiological responses associated with the Y5 receptor. Specifically, antagonists to the Y5 receptor could be used to treat obesity and diabetes by reducing appetite and food consumption, whereas agonists could be used for the treatment of anorexic conditions. The proteins are also useful for developing antibodies for detection of the protein.

Flowing from the foregoing are a number of other aspects of the invention, including (a) vectors, such as plasmids, comprising the receptor Y5 nucleic acid sequence that may further comprise additional regulatory elements, *e.g.*, promoters, (b) transformed cells that express the Y5 receptor, (c) nucleic acid probes, (d) antisense oligonucleotides, (e) agonists, (f) antagonists, and (g) transgenic mammals. Further aspects of the invention comprise methods for making and using the foregoing compounds and compositions.

The foregoing merely summarizes certain aspects of the present invention and is not intended, nor should it be construed, to limit the invention in any manner. All patents and other publications recited herein are hereby incorporated by reference in their entirety.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 displays the competition curves of various peptides for [125 I]PYY to Y5 receptor membranes transiently expressed in COS-7 cells.

Figure 2 displays saturation curves for specific binding of [125 I]PYY to Y5 receptor
5 membranes transiently expressed in COS-7 cells.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention comprises, in part, a novel NPY/PYY receptor protein, the Y5 receptor. Particularly preferred embodiments of the Y5 receptor are those having an amino acid sequence substantially the same as SEQ ID NOs 2, 4, or 6. As used herein, reference to the Y5
10 receptor is meant as a reference to any protein having an amino acid sequence substantially the same as SEQ ID NOs 2, 4, or 6. The present invention also comprises the nucleic acid sequence encoding the Y5 protein, which nucleic acid sequence is substantially the same as SEQ ID NOs 1, 3, or 5. Receptors SEQ ID NOs 2 and SEQ ID NO 4 are rat Y5 receptors and appear to be allelic variations, with SEQ ID NO 4 the most commonly occurring and, therefore, the
15 preferred embodiment of the rat Y5 receptor of this invention. SEQ ID NO 6 is the human Y5 receptor and its preferred embodiment.

As used herein, a protein "having an amino acid sequence substantially the same as SEQ ID NO x" (where "x" is the number of one of the protein sequences recited in the Sequence Listing) means a protein whose amino acid sequence is the same as SEQ ID NO x or differs
20 only in a way such that $IC_{50}[(3-36)NPY]$, $IC_{50}[(Leu^{31}Pro^{34})NPY]$, and $IC_{50}[(Leu^{31}Pro^{34})(3-36)NPY]$ as determined according to the method detailed in Example 4, *infra*, are less than or equal to 30 nM. The NPY fragments (3-36)NPY, (Leu³¹Pro³⁴)NPY and (Leu³¹Pro³⁴)(3-36)NPY induce a feeding response. Those skilled in the art will appreciate that conservative substitutions of amino acids can be made without significantly diminishing the protein's affinity
25 for NPY, PYY, and fragments and analogs thereof. Other substitutions may be made that increase the protein's affinity for these compounds. Making and identifying such proteins is a routine matter given the teachings herein, and can be accomplished, for example, by altering the nucleic acid sequence encoding the protein (as disclosed herein), inserting it into a vector, transforming a cell, expressing the nucleic acid sequence, and measuring the binding affinity of
30 the resulting protein, all as taught herein.

As used herein the term "a molecule having a nucleotide sequence substantially the same as SEQ ID NO y" (wherein "y" is the number of one of the protein-encoding nucleotide sequences listed in the Sequence Listing) means a nucleic acid encoding a protein "having an amino acid sequence substantially the same as SEQ ID NO y+1" (wherein "y+1" is the number of the amino acid sequence for which nucleotide sequence "y" codes) as defined above. This definition is intended to encompass natural allelic variations in the Y5 sequence. Cloned nucleic acid provided by the present invention may encode Y5 protein of any species of origin, including (but not limited to), for example, mouse, rat, rabbit, cat, dog, primate, and human. Preferably the nucleic acid provided by the invention encodes Y5 receptors of mammalian, and most preferably, rat or human origin.

The invention also includes nucleotide sequences encoding chimeric proteins comprised of parts of the Y5 receptor and parts of other related seven-transmembrane receptors.

The 6B clone (SEQ ID NO 1) (*see* Example 2, *infra*) has a 2.4 kb cDNA insert with a open reading frame from nucleotide 248 to 1582 that encodes a 445 amino acid protein (SEQ ID NO 2). Hydrophobicity plot analysis using PEPLOT of GCG shows that the Y5 receptor has seven transmembrane-like domains, indicating it might be a G-protein-coupled receptor. Unlike other known subtypes of NPY receptor family, the third intracellular loop of the Y5 receptor is unusually long. Another novel feature of the Y5 peptide sequence is that it has a much shorter C-terminal tail sequence than other known members of the NPY receptor family. It is also important to note that the Y5 sequence shows only 30-33% amino acid sequence identity to other NPY receptors.

Nucleic acid hybridization probes provided by the invention are DNAs consisting essentially of the nucleotide sequences complementary to any sequence depicted in SEQ ID NOs 1, 3, and 5 that is effective in nucleic acid hybridization. Nucleic acid probes are useful for detecting Y5 gene expression in cells and tissues using techniques well-known in the art, including, but not limited to, Northern blot hybridization, *in situ* hybridization, and Southern hybridization to reverse transcriptase - polymerase chain reaction product DNAs. The probes provided by the present invention, including oligonucleotide probes derived therefrom, are also useful for Southern hybridization of mammalian, preferably human, genomic DNA for screening for restriction fragment length polymorphism (RFLP) associated with certain genetic disorders. As used herein, the term complementary means a nucleic acid having a sequence that is sufficiently complementary in the Watson-Crick sense to a target nucleic acid to bind to the

target under physiological conditions or experimental conditions those skilled in the art routinely use when employing probes.

Receptor Y5 binds various fragments and analogs of NPY and PYY with affinities different from that of the known receptors. The rank order of binding affinity of receptor Y5 was found to be:

NPY = (LP)NPY = PYY = (3-36)NPY = (LP)(3-36)NPY > (10-36)NPY > (18-36)NPY

Table 1, *infra*, presents a more detailed affinity profile of the Y5 receptor for NPY, PYY, and various fragments thereof. As used herein, a protein having substantially the same affinity profile as the Y5 receptor means a protein in which the IC₅₀ of each of the peptides listed in Table 1, *infra*, is no more than an order of magnitude greater than those listed in Table 1 for each of the respective peptides as measured according to the methods described in Example 4. Importantly, the NPY analogs/fragments that induce feeding, such as (LP)(3-36)NPY, do not bind to the previously identified NPY/PYY receptors with affinities consistent with the feeding response.

The production of proteins such as receptor Y5 from cloned genes by genetic engineering means is well known in this art. The discussion which follows is accordingly intended as an overview of this field, and is not intended to reflect the full state of the art.

DNA which encodes receptor Y5 may be obtained, in view of the instant disclosure, by chemical synthesis, by screening reverse transcripts of mRNA from appropriate cells or cell line cultures, by screening genomic libraries from appropriate cells, or by combinations of these procedures, as illustrated below. Screening of mRNA or genomic DNA may be carried out with oligonucleotide probes generated from the Y5 gene sequence information provided herein. Probes may be labeled with a detectable group such as a fluorescent group, a radioactive atom or a chemiluminescent group in accordance with known procedures and used in conventional hybridization assays, as described in greater detail in the Examples below. In the alternative, the Y5 gene sequence may be obtained by use of the polymerase chain reaction (PCR) procedure, with the PCR oligonucleotide primers being produced from the Y5 gene sequence provided herein. See U.S. Patent Nos. 4,683,195 to Mullis *et al.* and 4,683,202 to Mullis.

Receptor Y5 may be synthesized in host cells transformed with a recombinant expression construct comprising a nucleic acid encoding the receptor Y5. Such a recombinant expression construct can also be comprised of a vector that is a replicable DNA construct. Vectors are used herein either to amplify DNA encoding Y5 and/or to express DNA which

encodes Y5. For the purposes of this invention, a recombinant expression construct is a replicable DNA construct in which a DNA sequence encoding Y5 is operably linked to suitable control sequences capable of effecting the expression of Y5 in a suitable host. The need for such control sequences will vary depending upon the host selected and the transformation method chosen. Generally, control sequences include a transcriptional promoter, an optional operator sequence to control transcription, a sequence encoding suitable mRNA ribosomal binding sites, and sequences which control the termination of transcription and translation. Amplification vectors do not require expression control domains. All that is needed is the ability to replicate in a host, usually conferred by an origin of replication, and a selection gene to facilitate recognition of transformants. See, Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual* (2nd Edition, Cold Spring Harbor Press, New York, 1989).

Vectors useful for practicing the present invention include plasmids, viruses (including phage), retroviruses, and integratable DNA fragments (*i.e.*, fragments integratable into the host genome by homologous recombination). The vector replicates and functions independently of the host genome, or may, in some instances, integrate into the genome itself. Suitable vectors will contain replicon and control sequences which are derived from species compatible with the intended expression host. The vectors may be self-replicating. Suitable vectors for the purposes of the present invention include pBluescript, pcDNA3, and, for insect cells, baculovirus. A preferred vector is the plasmid pcDNA3 (Invitrogen).

Construction of suitable vectors containing the desired coding and control sequences employs standard ligation and restriction techniques that are well understood in the art. Isolated plasmids, DNA sequences, or synthesized oligonucleotides are cleaved, tailored, and relegated in the form desired.

Site-specific DNA cleavage is performed by treating with the suitable restriction enzyme (or enzymes) under conditions that are generally understood in the art, and the particulars of which are specified by the manufacturer of these commercially available restriction enzymes. See, *e.g.*, *New England Biolabs, Product Catalog*. In general, about 1 μ g of plasmid or DNA sequence is cleaved by one unit of enzyme in about 20 μ l of buffer solution. Often excess of restriction enzyme is used to ensure complete digestion of the DNA substrate. Incubation times of about one hour to two hours at about 37°C are workable, although variations are tolerable. After each incubation, protein is removed by extraction with phenol/chloroform, and may be followed by ether extraction. The nucleic acid may be recovered from aqueous fractions by

precipitation with ethanol. If desired, size separation of the cleaved fragments may be performed by polyacrylamide gel or agarose gel electrophoresis using standard techniques. A general description of size separations is found in *Methods in Enzymology* 65, 499-560 (1980).

Transformed host cells are cells which have been transformed or transfected with recombinant expression constructs made using recombinant DNA techniques and comprising mammalian Y5-encoding sequences. Preferred host cells for transient transfection are COS-7 cells. Transformed host cells may ordinarily express Y5, but host cells transformed for purposes of cloning or amplifying nucleic acid hybridization probe DNA need not express the receptor. When expressed, the mammalian Y5 protein will typically be located in the host cell membrane. See, Sambrook *et al.*, *ibid.*

Cultures of cells derived from multicellular organisms are desirable hosts for recombinant Y5 protein synthesis. In principal, any higher eukaryotic cell culture is workable, whether from vertebrate or invertebrate culture. However, mammalian cells are preferred, as illustrated in the Examples. Propagation of such cells in cell culture has become a routine procedure. See Tissue Culture (Academic Press, Kruse & Patterson, Eds., 1973). Examples of useful host cell lines are bacteria cells, insect cells, yeast cells, human 293 cells, VERO and HeLa cells, LMTK⁻ cells, and WI138, BHK, COS-7, CV, and MDCK cell lines. Human 293 cells are preferred.

The invention provides homogeneous compositions of mammalian Y5 produced by transformed eukaryotic cells as provided herein. Such homogeneous compositions are intended to be comprised of mammalian Y5 protein that comprises at least 90% of the protein in such homogenous composition. The invention also provides membrane preparation from cells expressing Y5 as the result of transformation with a recombinant expression construct, as described here.

Mammalian Y5 protein made from cloned genes in accordance with the present invention may be used for screening compounds for Y5 agonist or antagonist activity, or for determining the amount of a Y5 agonist or antagonist drug in a solution (e.g., blood plasma or serum). For example, host cells may be transformed with a recombinant expression construct of the present invention, Y5 protein expressed in those host cells, the cells lysed, and the membranes from those cells used to screen compounds for Y5 binding activity. Competitive binding assays in which such procedures may be carried out are well known in the art. By selection of host cells which do not ordinarily express Y5, pure or crude preparations of

membranes containing Y5 can be obtained. Further, Y5 agonists and antagonists can be identified by transforming host cells with a recombinant expression construct as provided by the present invention. Membranes obtained from such cells (and membranes of intact cells) can be used in binding studies wherein the drug dissociation activity is monitored.

5 It is known that the neurotransmitter NPY is a regulator of appetite. As shown herein, the various NPY analogs/fragments that induce feeding, such as (LP)(3-36)NPY, bind with a high affinity to the Y5 receptor. Conversely, the NPY analogs/fragments that bind to the Y5 receptor with a lower affinity, such as (20-36)NPY, do not elicit feeding. It is therefore evident that by contacting the Y5 receptor with agonists and antagonists, feeding can be modulated. 10 Accordingly, antagonists to the Y5 receptor, identified by the methods described herein, can be used to reduce appetite and hence treat obesity, diabetes and hyperlipidemia, and, conversely, agonists to the Y5 receptor can be used to treat conditions such as anorexia.

This invention provides a pharmaceutical composition comprising an effective amount of a agonist or antagonist drug identified by the method described herein and a 15 pharmaceutically acceptable carrier. Such drugs and carrier can be administered by various routes, for example oral, subcutaneous, intramuscular, intravenous or intracerebral. The preferred route of administration would be oral at daily doses of about 0.01-100 mg/kg.

This invention provides a method of treating obesity, diabetes or hyperlipidemia, wherein the abnormality is improved by reducing the activity of Y5 receptor or blocking the 20 binding of ligands to a Y5 receptor, which method comprises administering an effective amount of the antagonist-containing pharmaceutical composition described above to suppress the subject's appetite. Similarly, the invention also provides methods for treating diseases and conditions resulting from underfeeding and/or a loss of appetite, which method comprises administering an effective amount of an agonist-containing pharmaceutical composition 25 described above to stimulate the subject's appetite.

The recombinant expression constructs of the present invention are useful in molecular biology to transform cells which do not ordinarily express Y5 to thereafter express this receptor.

Such cells are useful as intermediates for making cell membrane preparations useful for receptor binding assays, which are in turn useful for drug screening. Drugs identified from such 30 receptor assays can be used for the treatment of obesity, diabetes or anorexia.

The recombinant expression constructs of the present invention are also useful in gene therapy. Cloned genes of the present invention, or fragments thereof, may also be used in gene

therapy carried out by homologous recombination or site-directed mutagenesis. *See generally* Thomas & Capecchi, *Cell* **51**, 503-512 (1987); Bertling, *Bioscience Reports* **7**, 107-112 (1987); Smithies *et al.*, *Nature* **317**, 230-234 (1985).

Oligonucleotides of the present invention are useful as diagnostic tools for probing Y5 gene expression in tissues. For example, tissues are probed *in situ* with oligonucleotide probes carrying detectable groups by conventional autoradiographic techniques, as explained in greater detail in the Examples below, to investigate native expression of this receptor or pathological conditions relating thereto. Further, chromosomes can be probed to investigate the presence or absence of the Y5 gene, and potential pathological conditions related thereto, as also illustrated by the Examples below. Probes according to the invention should generally be at least about 15 nucleotides in length to prevent binding to random sequences, but, under the appropriate circumstances may be smaller.

The invention also provides antibodies that are immunologically reactive to a mammalian Y5, preferably rat or human Y5. The antibodies provided by the invention are raised in animals by inoculation with cells that express a mammalian Y5 or epitopes thereof, using methods well known in the art. Animals that are used for such inoculations include individuals from species comprising cows, sheep, pigs, mice, rats, rabbits, hamsters, goats and primates. Preferred animals for inoculation are rodents (including mice, rats, hamsters) and rabbits. The most preferred animal is the mouse.

Cells that can be used for such inoculations, or for any of the other means used in the invention, include any cell line which naturally expresses a mammalian Y5, or any cell or cell line that expresses a mammalian Y5 or any epitope thereof as a result of molecular or genetic engineering, or that has been treated to increase the expression of a mammalian Y5 by physical, biochemical or genetic means. Preferred cells are human cells, most preferably HEK 293 and BHK cells that have been transformed with a recombinant expression construct comprising a nucleic acid encoding a mammalian Y5, preferably a rat or human Y5, and that express the mammalian Y5 gene product.

The present invention provides monoclonal antibodies that are immunologically reactive with an epitope of mammalian Y5 or fragment thereof and that is present on the surface of mammalian cells, preferably human or mouse cells. These antibodies are made using methods and techniques well known to those of skill in the art.

Monoclonal antibodies provided by the present invention are produced by hybridoma cell lines, that are also provided by the invention and that are made by methods well known in the art. Hybridoma cell lines are made by fusing individual cells of a myeloma cell line with spleen cells derived from animals immunized with cells expressing the Y5 receptor, preferably rat or human cells, as described above. The myeloma cell lines used in the invention include lines derived from myelomas of mice, rats, hamsters, primates and humans. Preferred myeloma cell lines are from mouse. The animals from whom spleens are obtained after immunization are rats, mice and hamsters, preferably mice, most preferably Balb/c mice. Spleen cells and myeloma cells are fused using a number of methods well known in the art, including but not limited to incubation with inactivated Sendai virus and incubation in the presence of polyethylene glycol (PEG). The most preferred method for cell fusion is incubation in the presence of a solution of 45% (w/v) PEG-1450. Monoclonal antibodies produced by hybridoma cell lines can be harvested from cell culture supernatant fluids from *in vitro* cell growth; alternatively, hybridoma cells can be injected subcutaneously and/or into the peritoneal cavity of an animal, most preferably a mouse, and the monoclonal antibodies obtained from blood and/or ascites fluid.

Monoclonal antibodies provided by the present invention are also produced by recombinant genetic methods well known to those of skill in the art, and the present invention encompasses antibodies made by such methods that are immunologically reactive with an epitope of a mammalian Y5.

The present invention encompasses fragments of the antibody that are immunologically reactive with an epitope of a mammalian Y5. Such fragments are produced by any number of methods, including but not limited to proteolytic cleavage, chemical synthesis or preparation of such fragments by means of genetic engineering technology. The present invention also encompasses single-chain antibodies that are immunologically reactive with an epitope of a mammalian Y5 made by methods known to those of skill in the art.

The present invention also encompasses an epitope of a mammalian Y5 that is comprised of sequences and/or a conformation of sequences present in the mammalian Y5 molecule. This epitope may be naturally occurring, or may be the result of proteolytic cleavage of the mammalian Y5 molecule and isolation of an epitope-containing peptide or may be obtained by synthesis of an epitope-containing peptide using methods well known to those skilled in the art. The present invention also encompasses epitope peptides produced as a result

of genetic engineering technology and synthesized by genetically engineered prokaryotic or eukaryotic cells.

The invention also includes chimeric antibodies, comprised of light chain and heavy chain peptides immunologically reactive to an epitope that is a mammalian Y5. The chimeric antibodies embodied in the present invention include those that are derived from naturally occurring antibodies as well as chimeric antibodies made by means of genetic engineering technology well known to those of skill in the art.

Also provided by the present invention are non-human transgenic animals grown from germ cells transformed with the Y5 nucleic acid sequence according to the invention and that express the Y5 receptor according to the invention and offspring and descendants thereof. Also provided are transgenic non-human mammals comprising a homologous recombination knockout of the native Y5 receptor, as well as transgenic non-human mammals grown from germ cells transformed with nucleic acid antisense to the Y5 nucleic acid of the invention and offspring and descendants thereof. Further included as part of the present invention are transgenic animals which the native Y5 receptor has been replaced with the human homolog. Of course, offspring and descendants of all of the foregoing transgenic animals are also encompassed by the invention.

Transgenic animals according to the invention can be made using well known techniques with the nucleic acids disclosed herein. *E.g.*, Leder et al., U.S. Patent Nos. 4,736,866 and 5,175,383; Hogan et al., *Manipulating the Mouse Embryo, A Laboratory Manual* (Cold Spring Harbor Laboratory (1986)); Capecchi, *Science* 244, 1288 (1989); Zimmer and Gruss, *Nature* 338, 150 (1989); Kuhn et al., *Science* 269, 1427 (1995); Katsuki et al., *Science* 241, 593 (1988); Hasty et al., *Nature* 350, 243 (1991); Stacey et al., *Mol. Cell Biol.* 14, 1009 (1994); Hanks et al., *Science* 269, 679 (1995); and Marx, *Science* 269, 636 (1995). Such transgenic animals are useful for screening for and determining the physiological effects of Y5 receptor agonists and antagonist. Consequently, such transgenic animals are useful for developing drugs to regulate physiological activities in which NPY and/or PYY participate.

The following Examples are provided for illustrative purposes only and are not intended, nor should they be construed, as limiting the invention in any manner.

EXAMPLES

Example 1

Isolation and Sequencing of Rat Y5 Receptor

Isolation of rat hypothalamus mRNA and construction of cDNA library

5 Expression cloning strategy was used to clone novel NPY receptor in rat hypothalamus cDNA library. RNA was obtained from 9 frozen rat hypothalami weighing a total of 0.87 grams. Poly(A) RNA was isolated directly from the tissue using the Promega PolyATtract System 1000 kit (Promega, Madison, WI). The hypothalami were homogenized in 4 mL of 4M guanidine thiocyanate-25mM sodium citrate, pH 7.1-2% β -mercaptoethanol using a Polytron at
10 full-speed for approximately 1 minute. To the homogenized tissue 8 mL of 4M guanidine thiocyanate-25mM sodium citrate, pH 7.1-1% β -mercaptoethanol which had been preheated to 70°C was added. After mixing thoroughly, 870 pmol biotinylated oligo(dT) was added; the mixture was incubated at 70°C for 5 minutes. The homogenate was subjected to centrifugation at 12000 x g for 10 minutes at room temperature; the homogenate was transferred to a clean
15 tube and 10.44 mL Streptavidin MAGNESPHERE[®] Paramagnetic Particles (SA-PMPs) which had been prepared as per the published protocol was added. (Promega Corp. published protocol TM 228; Promega Corporation, Madison, WI). The homogenate and SA-PMPs were incubated together for 2 minutes at room temperature after which the homogenate was decanted while the SA-PMP-biotinylated oligo(dT)-hypothalamic poly(A) RNA complex was retained in the tube
20 by a magnetic stand. The complex was washed as per the protocol, after which the RNA was precipitated and resuspended in water. 25 micrograms of this poly(A) RNA was used by Invitrogen (Invitrogen Corporation, San Diego, CA) to prepare a cDNA expression library. The protocols used by Invitrogen to prepare the cDNA library are essentially based upon the procedures of Okayama and Berg (*Molec. Cell. Biol.* 2, 161 (1982)) and Gubler and Hoffman
25 (*Gene* 25, 263 (1983)) (Invitrogen Corporation publications 130813sa and 130928sa). An oligo(dT) anchor primer was used for reverse transcription, and the library was cloned unidirectionally into pcDNA3 vector which contains a CMV promoter for eukaryotic expression. The cDNA library had 5.3×10^5 primary recombinants with an average insert size of 2.59 kb.

Isolation of a novel Y5 receptor cDNA clone

The rat hypothalamus cDNA library was plated on the LB/Ampicillin plates in pools of 1,000 independent colonies. The plates were incubated at 37°C for about 20 hours and the bacteria from each plate were scraped in 4-5 ml LB/Ampicillin media. Two ml of the bacteria samples were used for plasmid preparation and one ml of each pool was stored at -80°C in 15% glycerol.

COS-7 cells were grown in Dulbecco's Modified Eagle Medium (DMEM, GIBCO 11965-092), 10% fetal bovine serum (GIBCO 16000-028), and 1 x antibiotic/antimycotic solution (GIBCO 15240-039) (Gaithersburg, MD). Cells were trypsinized and split at 50 to 70% confluency.

DNA from 1300 pools was transfected into COS-7 cells for [¹²⁵I]PYY binding assays. Twenty four hours before transfection, cells were plated into flaskette chambers (Nunc, Inc. 177453, Naperville, IL) at 3x10⁵ cells/flaskette (equivalent to 3x10⁴ cells/cm²). Two µg of plasmid DNA from each pool was transfected into the cells using 10 µl of Lipofectamine (GIBCO 18324-012) according to the manufacture's protocol. Forty eight hours after transfection, the [¹²⁵I]PYY binding assay was performed in the flaskette chamber. The cells were treated with 2 ml total binding buffer: 10 mM HEPES, 5 mM KCl, 1.2 mM KH₂PO₄, 2.5 mM CaCl₂, 1.2 mM MgSO₄, 150 mM NaCl, 25 mM NaHCO₃, 10 mg/ml bovine serum albumin, 0.5 mg/ml bacitracin and 0.4 mg/ml soybean trypsin inhibitor at room temperature for 15 minutes. The cells were then incubated with 100 pM porcine [¹²⁵I]PYY (Amersham (Arlington Heights, IL), Specific Activity 4000Ci/mmol) in the total binding buffer for 90 minutes at room temperature. After binding, the cells were washed three times with ice-cold total binding buffer without ligand and one time with cold phosphate buffered saline (PBS). Cells were fixed with 1% cold glutaraldehyde in PBS for 15 minutes, washed once with cold PBS/0.5 M Tris, pH 7.5 and incubated in PBS/0.5 M Tris, pH 7.5 for 15 minutes at 4°C. After washing one more time with cold PBS, the slides were dipped in 0.5% gelatin at 42°C and dried under vacuum. The dried slides were dipped in 50% photographic emulsion (Kodak (Rochester, NY) NTB2) at 42°C and exposed in the darkbox for four days at 4°C. After four days of exposure, the darkbox was moved to room temperature for one hour and slides were developed in developer D-19 (Kodak) for three minutes at 15°C and fixed in fixer (Kodak) for three minutes at 15°C, washed in water and air dried. Cells were stained with Diff-Quik stain

set (Baxter, McGaw Park, IL) and air dried. Slides were dipped into xylenes and mounted with DPX mountant (Electron Microscopy Science, Fort Washington, PA). Positive cells were identified using dark field microscopy.

Twenty one positive pools were identified. Since the hypothalamus expresses different subtypes of NPY receptors including Y1 and Y2 receptors, we analyzed all the positive pools for Y1, Y2 and Y4/PP receptors by PCR. Of the 21 positive pools tested as described above, 12 pools contained Y1, 4 pools contained Y2 and none contained Y4/PP. Five pools (Y217, Y555, Y589, Y861 and Y1139) were negative by PCR analysis. The pool Y217 was subdivided in 24 subpools of 200 colonies, then 50 colonies, and finally a single clone, the Y217.24.13.6B clone (6B), was isolated.

DNA and peptide sequences analysis

Plasmid DNA was sequenced by Lark Technologies Inc. (Houston, Texas) and Biotechnology Resource Laboratory of Yale University (New Haven, CT) using Sequenase Kit (US Biochemical, Cleveland, OH) or Applied Biosystems' automatic sequencer system (model 373A). The peptide sequence was deduced from the long open-reading-frame of the nucleotide sequence. DNA and peptide sequences were analyzed using the GCG program (Genetics Computer Group, Madison, WI). The results are embodied in SEQ ID NO 1 (the nucleic acid sequence) and SEQ ID NO 2 (the amino acid sequence).

Example 2

Localization of Rat Y5 Receptor in Brain and Other Tissues

Northern Blot

To study the expression level of the Y5 receptor in the rat brain and other tissues, we did Northern blot analysis using the 6B 2.4 kb probe. A rat multiple tissue Northern blot (Clontech Laboratories, Palo Alto, CA) was hybridized to the ³²P-labeled rat 6B probe. The blot contains 2 µg of poly A⁺ RNA per lane from rat heart, brain, spleen, lung, liver, skeletal muscle, kidney, and testis. Hybridization was carried out in 1x hybridization solution containing 6X SSC (0.9 M NaCl, 0.09 M Na Citrate, pH 7.0), 5x Denhardt's solution (0.1% polyvinylpyrrolidone, 0.1% ficoll type 400, 0.1% bovine serum albumin), 100 mg/ml sheared, and denatured salmon sperm DNA at 65°C. The filter was washed at 65°C in 0.1X SSC, 0.1% SDS and exposed to Kodak XAR 5 film with two intensifying screens. A single 2.6 kb band was detected in the brain after

overnight exposure of the blot. No bands were found from other tissues (heart, spleen, lung, liver, skeletal muscle, kidney and testis) in the Clontech multiple tissue Northern blot, even after six days of exposure.

We tested 6B expression in more rat tissues and different regions of brain. mRNA was isolated from rat whole brain, cortex, hypothalamus, hippocampus, olfactory bulb, spleen, stomach, kidney, small intestine, adrenal and pancreas using Fast Track Isolation Kit (Invitrogen). Ten µg of mRNA from different brain regions and multiple tissues were run on a denaturing formaldehyde 1% agarose gel, transferred to a Nytran membrane (Schleicher and Schuell) and hybridized with ³²P-labeled 6B 2.4 kb probe and washed at high stringency. After overnight hybridization, the filter was washed at high stringency and exposed to X-ray film with intensifying screens. The 6B receptor mRNA was detectable in the brain regions examined after one day exposure, but no signal was observed from other tissues, even after a week exposure with double intensifying screens.

Example 3

Isolation of Two Isoforms of the Rat Y5 Receptor

Plasmid DNA from pools Y555, Y589, and Y861 described in Example 1 were hybridized to the Y5 probe at high stringency. A single positive clone was isolated from the Y555 pool and sequenced as described in Example 1. Compared to the 6B DNA sequence, the Y555 sequence (SEQ ID NO 4 has a 123 bp insert sequence located at the 5'-untranslated region between nucleotides 239 and 240 of Y5 clone. The coding region of the clones Y555, Y589, and Y861 has the same sequence as clone 6B, except for one nucleotide substitution (C to T) at position 430 of the 6B clone. The nucleotide substitution changes the amino acid proline to leucine in the first transmembrane domain. The corresponding amino acid sequence is given by SEQ ID NO 4.

The different isoforms of the receptor could be the allelic variants of the same gene. To test this hypothesis, we analyzed genomic DNA from 16 rats. The genomic DNA from each animal was used as template for PCR analysis. A 314 bp DNA fragment that contains the site of the nucleotide variation was amplified and sequenced. Of the 16 DNA samples tested, 14 samples had a T at position 430 and 2 samples had a C. This result strongly suggests that the amino acid variation is an allelic variant.

Example 4

Pharmacological Characterization of the Novel Rat NPY Receptors

Transient Transfection

Monkey kidney cells (COS-7) were maintained in T-175 cm² flasks (NUNC) at 37°C with 5% CO₂ in a humidified atmosphere. Cells were grown in Dulbecco's Modified Eagle Medium (DMEM) supplemented with 2 mM glutamine, 10% fetal bovine serum, 1 mM sodium pyruvate, and antibiotic/antimycotic. Cells at 70% confluency were transfected with Y5 DNA using the Lipofectamine method (GIBCO-BRL). 15 µg DNA and 90 µl Lipofectamine were added to each flask. Media was completely replaced 24 hours post transfection, and membranes were harvested 24 hours later.

Stable Expression of the Rat NPY Y5 receptor (clone Y861)

A strain of the human embryonic kidney cell line 293 adapted to grow in suspension (293S) was used for these experiments. Approximately 1×10^6 cells were seeded onto a 10-cm dish 24 hours prior to transfection. The rat NPY Y5 cDNA (Y861), subcloned in the eukaryotic expression vector pcDNA3 (Invitrogen, Carlsbad, CA) was first linearized with *NotI* and purified using a Wizard PCR Prep kit (Promega). In preparation for transfection, 15 µg of the linearized DNA were added to 500 µl of DMEM cell culture media, and 30 µl of Lipofectamine (Life Sciences) were added into a separate 500 µl aliquot of DMEM. These two solutions were mixed together and incubated for 20 minutes at room temperature and the resulting DNA/lipid complexes were then slowly added to the cells (which had been previously rinsed once with serum-free DMEM) and covered with a total volume of 10 ml. Cells were then transferred to a humidified 10% CO₂ incubator and left for 4 hours at 37°C, at which time the media was replaced with DMEM supplemented with 8% FBS. After 16 hours, cells were trypsinized and split at a 1:15 ratio into 10-cm dishes containing DMEM/8% FBS in the presence of 700 µg/ml of G418 (selection media). When discrete colonies became apparent (after approximately 10 days), cells were pooled and carried through 2 additional passages in selection media. Cells were then trypsinized and diluted in preparation for cloning by limited dilution (CBLD), such that an average of one cell was seeded in each well of a 96-well microtiter culture plate, and was inspected periodically for the subsequent 2 to 3 weeks. After 21 days in culture under selection conditions, those wells containing single colonies were selected and transferred to 24-well culture plates following trypsinization. Each of these clones was propagated until

sufficient quantities were available for testing [125 I]PYY binding activity, from which one particular clone designated E7 was selected on the basis of its high level of binding activity.

Stable Expression of the human NPY Y5 receptor

293 cells were plated onto a T75 flask one day prior to transfection such that they were
5 50-70% confluent when used for the experiment. The human NPY Y5 intronless genomic clone HG.PCR15, containing the full length open reading frame encoding the receptor, was first linearized with *Not* I and purified using a Wizard PCR Prep kit (Promega). For each transfection, 8 μ g of linearized DNA were added to 1.25 ml of Optimem culture media (Life Sciences) and 37 μ l of Transfectam (Promega) were added to 1.25 ml of Optimem. These two
10 solutions were then mixed together and added to cells previously washed once with Optimem. After an incubation period of 5 hours, the DNA/Transfectam mixture was removed, cells were washed with PBS and fed with DMEM supplemented with 10% FBS. Cells were left intact for two days, and then switched to selection media (DMEM 10% FBS containing 350 μ g/ml of G418) for 5-10 days followed by CBLD as described above. The individual clone 293.hy5.sb.8
15 was selected on the basis of its high level of [125 I]PYY binding activity, using the intact cell binding protocol from above.

Membrane Preparation

The media was removed from each flask of transfected cells, and the cells were washed twice with 20 ml ice-cold phosphate buffered saline. The cells were scraped from the flask in 5
20 ml of Tris buffer (20 mM Tris-HCl and 5 mM EDTA, pH 7.7), and then transferred to a centrifuge tube. Each flask was washed with an additional 5 ml of Tris buffer and combined in the centrifuge tube. The cells were polytroned for 2 x 10 seconds (12 mm probe, 7000-8000 rpm) and centrifuged 5 minutes (Centra 7R, International Equipment Co., Needham Heights, MA) at 800 rpm and 4°C. The supernatant was then transferred to a clean centrifuge tube and
25 was centrifuged at 30,000 x g for 30 minutes and 4°C. The supernatant was removed and the pellet was stored at -80°C. Protein concentration was measured using the Bio-Rad kit pursuant to the standard manufacturer's protocol (Biorad Laboratories, Hercules, CA) with bovine IgG as the standard.

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Stable Expression of the human NPY Y5 receptor

293 cells were plated onto a T75 flask one day prior to transfection such that they were
5 50-70% confluent when used for the experiment. The human NPY Y5 intronless genomic clone HG.PCR15, containing the full length open reading frame encoding the receptor, was first linearized with *Not* I and purified using a Wizard PCR Prep kit (Promega). For each transfection, 8 µg of linearized DNA were added to 1.25 ml of Optimem culture media (Life Sciences) and 37 µl of Transfectam (Promega) were added to 1.25 ml of Optimem. These two
10 solutions were then mixed together and added to cells previously washed once with Optimem. After an incubation period of 5 hours, the DNA/Transfectam mixture was removed, cells were washed with PBS and fed with DMEM supplemented with 10% FBS. Cells were left intact for two days, and then switched to selection media (DMEM 10% FBS containing 350 µg/ml of G418) for 5-10 days followed by CBLD as described above. The individual clone 293.hy5.sb.8
15 was selected on the basis of its high level of [¹²⁵I]PYY binding activity, using the intact cell binding protocol from above.

Membrane Preparation

The media was removed from each flask of transfected cells, and the cells were washed twice with 20 ml ice-cold phosphate buffered saline. The cells were scraped from the flask in 5
20 ml of Tris buffer (20 mM Tris-HCl and 5 mM EDTA, pH 7.7), and then transferred to a centrifuge tube. Each flask was washed with an additional 5 ml of Tris buffer and combined in the centrifuge tube. The cells were polytroned for 2 x 10 seconds (12 mm probe, 7000-8000 rpm) and centrifuged 5 minutes (Centra 7R, International Equipment Co., Needham Heights, MA) at 800 rpm and 4°C. The supernatant was then transferred to a clean centrifuge tube and
25 was centrifuged at 30,000 x g for 30 minutes and 4°C. The supernatant was removed and the pellet was stored at -80°C. Protein concentration was measured using the Bio-Rad kit pursuant to the standard manufacturer's protocol (Biorad Laboratories, Hercules, CA) with bovine IgG as the standard.

¹²⁵I]PYY Binding Assay for NPY Y5 Receptors

The binding assays were performed on GF/C Millipore (Bedford, MA) 96-well plates pretreated with 0.02% polyethylenimine (PEI) for at least 2 hours prior to use. The PEI was aspirated from the plates on a vacuum manifold immediately before the samples were added to the wells. All peptides, tissue and radioligand were diluted with binding buffer (25 mM Tris, 120 mM NaCl, 5 mM KCl, 1.2 mM KH₂PO₄, 2.5 mM CaCl₂, 1.2 mM MgSO₄, 0.1% BSA and 0.5 mg/ml bacitracin, pH 7.4). For competition assays, increasing concentrations of peptide were incubated with [¹²⁵I]PYY and tissue. In a final volume of 200 µl, samples consisted of: membrane protein (*i.e.*, 2.5-15 or 10-30 µg membrane protein for rat Y5 or human Y5, respectively); 75-100 pM [¹²⁵I]PYY NEN-DuPont (Boston, MA); peptide dilution or binding buffer. Nonspecific binding was defined by 1 µM PYY. NPY, PYY, (2-36)NPY, (10-36)NPY, (LP)(3-36)NPY and (32D-Trp)NPY were synthesized at Bayer Corp. (West Haven, CT). All other peptides were purchased from either Peninsula (Belmont, CA) or Bachem (Torrance, CA).

For saturation experiments, increasing concentrations of [¹²⁵I]PYY were incubated with membrane and 1 µM PYY. After a 2 hour incubation at room temperature with constant mixing, the samples were aspirated on a vacuum manifold. The wells were washed with three 200 µl aliquots of ice-cold binding buffer. The individual wells were punched into 12x75 mm plastic tubes, and counted on a Wallac (Gaithersburg, MD) gamma counter. Binding data were analyzed using the nonlinear regression curve-fitting program RS/1 (BBN Software Products Corp., Cambridge, MA).

Binding Assays for Rat Y2, Y1, and Y4/PP1 Receptors

The binding buffer for rat Y2 binding was Krebs/Ringer bicarbonate (Sigma K-4002, S-8875), pH 7.4, containing 0.01% bovine serum albumin (BSA - Sigma A-2153) and 0.005% bacitracin. 0.85-1 µg of protein and 25 pM [¹²⁵I]PYY are added to each well. Nonspecific binding is defined by 1 µM NPY.

The binding buffer for rat Y1 and rat Y4/PP1 binding consisted of 137 mM NaCl, 5.4 mM KCl, 0.44 mM KH₂PO₄, 1.26 mM CaCl₂, 0.81 mM MgSO₄, 20 mM HEPES, 1 mM dithiothreitol (DTT), 0.1% bacitracin, 100 mg/l streptomycin sulfate, 1 mg/l aprotinin, 10 mg/ml soybean trypsin inhibitor and 0.3% BSA, pH 7.4. For rat Y1 binding, ~5-15 µg of protein and 50 pM [¹²⁵I]PYY were added to each well, and nonspecific binding was defined by 1 µM NPY. For the rat Y4/PP1 binding assay, ~1-2 µg of protein and 50 pM rat [¹²⁵I]PP (NEN

DuPont, Boston, MA) were added to each well, and 1 μ M rat PP was used to define nonspecific binding.

In Vitro Functional Assay - Measurement of Forskolin-Stimulated Adenylate Cyclase

Rat Y5

5 (Reference: Gordon et al., *J. Neurochem.* 55, 506, 1990) Suspension cells stably expressing the Y5 receptor (approximately 400,000 per sample) were resuspended in serum-free DMEM containing 10mM HEPES (pH 7.4) and 1 mM isobutylmethylxanthine (IBMX). 1 μ M forskolin was added to the cells. The assay was stopped by transferring the samples into a boiling water bath for 3 minutes. After a 3 minute centrifugation at 14,000xg, an aliquot of
10 each sample was quantitated for cAMP levels by radioimmunoassay (NEN DuPont, MA).

Human Y5

Monolayer cells stably expressing the Y5 receptor were pre-rinsed with Wash buffer (pH 7.2: 137 mM NaCl, 2.7 mM KCl, 0.9 mM CaCl₂, 0.5 mM MgCl₂, 6.5 mM Na₂HPO₄, 1.5 mM KH₂PO₄). Cells were then incubated for 10 minutes at 37°C in Assay buffer (pH 7.4:
15 Wash buffer + 10 mM HEPES, 10 mg/ml BSA, 0.5 mg/ml bacitracin, 0.4 mg/ml soybean trypsin inhibitor). After addition of fresh buffer and 100 μ M IBMX, the cells were incubated for 10 minutes at 37°C. The reaction was started with the addition of peptide and 1-10 μ M forskolin. After a 20 minute incubation at 37°C, the reaction was terminated by discarding the
20 buffer and adding 65% ethanol to each well. The supernatant was then transferred to microfuge tubes and the extraction step was repeated once more. After evaporation of the ethanol from the samples, the amount of cAMP was assayed using by radioimmunoassay (NEN DuPont: Boston, MA).

In Vivo Pharmacology Procedures

Adult male Wistar rats were surgically implanted with a chronic intracerebral ventricular
25 (ICV) cannula (Plastic Products, Roanoke, Virginia) using a stereotaxic instrument. Several days after the surgery, 1-6 nmoles of each peptide (or saline) was injected into the lateral ventricle of 4-12 rats in a volume of 5-10 μ l. The quantity of rodent chow consumed in a 2 hour period was measured.

In Vitro and In Vivo Pharmacology Results

Figure 1 presents the competition curves of various peptides for [¹²⁵I]PYY binding to Y5 receptor membranes transiently expressed in COS-7 cells. Each point is the average value of triplicate determinations from a representative experiment. IC₅₀ values corresponding to 50% inhibition of specific binding were determined using nonlinear regression analysis. K_i values were calculated from the IC₅₀ values using the Cheng-Prusoff correction, such that $K_i = IC_{50} / (1 \pm (L/K_d))$, where L is the radioligand concentration and K_d is the dissociation constant. The results for transiently expressed Y5 clones are presented in Table 1, and Table 2 contains data for stably expressed Y5 clones.

Table 1

PEPTIDE	NPY Y5 BINDING AFFINITIES (K _i ± SEM, nM)			
	RAT Y555 Clone*	RAT 6B Clone	RAT Y861 Clone	HUMAN Clone
r/hNPY	0.53 ± 0.06	0.49 ± 0.03	0.50 ± 0.06	0.73 ± 0.09
rPYY	1.1 (1.2, 0.95)	0.48 ± 0.09	1.0 ± 0.13	1.3 ± 0.14
h(LP)PYY	2.5 ± 0.5	0.57 ± 0.01	1.8 ± 0.09	1.7 ± 0.3
r/h(LP)NPY	0.96 (1.0, 0.92)	0.31	0.55 ± 0.11	0.97 ± 0.36
p(LP)NPY	ND	0.64 ± 0.07	0.47	0.88 ± 0.11
r/h(2-36)NPY	0.81 (0.61, 1)	0.65	1.2 ± 0.07	1.2 ± 0.15
p(3-36)NPY	3.6 ± 0.4	1.9 ± 0.27	2.0 (1.8, 2.3)	10.4 ± 2.0
r/h(3-36)NPY	ND	0.49	2.1 (2.7, 1.6)	3.8 ± 0.48
r(3-36)PYY	6.2 ± 1.1	1.4 ± 0.10	4.2 ± 0.47	10 ± 3.4
r/h(10-36)NPY	35	4.9 (6.0, 3.8)	34 ± 2.8	110 (110, 109)
p(13-36)NPY	40 (38, 41)	7.7 (7.9, 7.5)	22 (25, 19)	56 ± 7
r(13-36)NPY	73	11 ± 1.0	86 ± 19	77 (89, 65)
p(18-36)NPY	303	194 ± 88	206 ± 61	618 ± 85
r/h(20-36)NPY	636	330 ± 31	587	>1000
r/h(22-36)NPY	>1000	>1000	>1000	>1000
r/h(26-36)NPY	>1000	>1000	>1000	>1000
(1-24)NPY	ND	>1000	>1000	>1000
BIBP3226	ND	>1000	>1000	>1000
hPP	ND	ND	4.0 ± 0.29	11 (15, 6.2)
rPP	ND	62	296 ± 47	436 (582, 290)

* IC₅₀ values (nM)

Table 2

Peptide	K _i Values (nM; Average \pm SEM)	
	293.hY5.sb.8	293S.Y861.2
rPYY	1.3 \pm 0.2	0.71 \pm 0.1
hPYY	1.1 \pm 0.2	1.06 \pm 0.2
(3-36)PYY	4.5 \pm 0.7	3.6 \pm 0.4
(13-36)PYY	24 \pm 2.0	29 \pm 4
h(LP)PYY	1.3 \pm 0.1	0.76 \pm 0.1
r/hNPY	0.79 \pm 0.1	0.86 \pm 0.07
p(LP)NPY	1.2 \pm 0.4	0.67 \pm 0.04
h/r(LP)NPY	0.89 \pm 0.1	0.67 \pm 0.04
(LP)(3-36)NPY	3.1 \pm 0.6	2.9 \pm 0.9
(2-36)NPY	1.4 \pm 0.03	0.83 \pm 0.1
h(3-36)NPY	3.5 \pm 0.4	1.4 \pm 0.4
(10-36)NPY	14 \pm 2.7	15 \pm 4.7
p(13-36)NPY	8.7 \pm 1.6	8.8 \pm 2.0
p(18-36)NPY	144 \pm 18	61 \pm 13
(20-36)NPY	429 \pm 133	108 \pm 16
(22-36)NPY	>900	>930
(26-36)NPY	>900	>930
(1-24)NPY	>900	>930
(32D-Trp)NPY	7.3 \pm 0.8	4.2 \pm 1.0
hPP	3.7 \pm 1.6	2.5 \pm 0.5
rPP	286 \pm 77	203 \pm 44

K_i values for various peptides for [¹²⁵I]PYY binding to the transiently expressed rat 6B, Y861 and Y555 receptor clones as well as the human Y5 receptor. The averages \pm standard error of the mean (SEM) represent values from at least three independent experiments. Two independent experiments are represented by the average, followed by the individual values in parentheses. Remaining values without SEM are from a single experiment. Peptide species in Table 1 (and Table 2, *infra*) are indicated with the following prefixes: r = rat, h = human, p = porcine, r/h = rat = human. ND = not determined.

The rank order of the affinities of the peptides tested is as follows:

NPY ~ PYY ~ (LP)PYY ~ (LP)NPY ~ (2-36)NPY ~ (3-36)PYY ~
 (LP)(3-36)NPY ~ (3-36)NPY > (32D-Trp)NPY > (10-36)NPY ~ (13-36)NPY >
 (18-36)NPY > (20-36)NPY > > (22-36)NPY, (26-36)NPY

In Table 3, the pharmacological profile of the standard peptides is expanded for the other cloned NPY receptors to further illustrate the novel nature of the Y5 receptor pharmacology. In addition, the *in vivo* feeding response of some of these peptides is listed for comparison. The data shown are representative of the average of at least two independent experiments, as described in the methods. Feeding of rats injected (ICV) with saline was < 3g/2hours.

Table 4 shows the EC₅₀ values for same standard peptides at the rat and human Y5 receptor.

C-terminal fragment (3-36)NPY binds preferentially to Y2 receptors, while (LP)NPY has lower affinity. Conversely, (LP)NPY has high affinity for the Y1 receptor, while (3-36)NPY and the C-terminal fragments are much weaker. When considering the rat Y4/PP1 receptor, rat PP has very high affinity as compared to NPY, PYY, (LP)NPY, and (13-36)NPY. In the *in vivo* feeding model, (LP)NPY, which has high affinity for Y1 and low affinity for Y2, and (3-36)NPY, which has a high affinity for Y2, but not Y1, all stimulate feeding in rats. Rat PP does not induce much feeding when administered to rats. This *in vivo* profile matches the *in vitro* pharmacological profile outlined in Table 2 for the Y5 receptor.

In addition, while (LP)(3-36)NPY (a custom peptide synthesized at Bayer) has weak affinity for Y1, Y2 and Y4/PP1, it stimulates feeding in rats. Importantly, (LP)(3-36)NPY has high affinity for the Y5 receptor (Table 2). These data are further evidence that the Y5 receptor is linked to feeding.

Table 3

IC ₅₀ VALUE (nM)					
PEPTIDE	Rat Y1 (clone)	Rat Y2 (clone)	Rat Y4/PP1 (clone)	Rat Y5 (Y861)	Feeding (g/2 h)
r/hNPY	0.13	0.24	> 1000	0.45	>5
rPYY	0.43	0.079	630	0.9	>5
h(Leu ³¹ Pro ³⁴)PYY	0.57	116	ND	2.0	>5

IC ₅₀ VALUE (nM)					
PEPTIDE	Rat Y1 (clone)	Rat Y2 (clone)	Rat Y4/PP1 (clone)	Rat Y5 (Y861)	Feeding (g/2 h)
p(Leu ³¹ Pro ³⁴) NPY	0.15	150	4.3	0.63	>5
r/h(2-36)NPY	47	0.50	>1000	1.3	>5
p(3-36)NPY	45	0.67	>1000	2.2	>5
r/h(Leu ³¹ Pro ³⁴) (3-36)NPY	44	154	20	3.4	>5
hPP	40	>1000	0.065	4.9	>5
(32DT _{Trp})NPY	> 1000	26	ND	7.0	ND
r/h(10-36)NPY	148	0.42	>1000	34	<3
rPP	843	>1000	0.071	325	<3
p(18-36)NPY	287	0.34	159	326	<3
(20-36)NPY	435	0.64	ND	638	<3
(22-36)NPY	>1000	0.89	ND	>1000	<3
(26-36)NPY	>1000	84	ND	>1000	<3
(1-24)NPY	>1000	>1000	ND	>1000	<3

The pharmacological profile for the 6B (and Y861 and Y555) receptor clones is distinct from Y1 receptors (where PYY~NPY~(LP)NPY > (3-36)NPY > (13-36)NPY ~ (18-36)NPY > (LP)(3-36)NPY), as well as Y2 receptors (where PYY~NPY~(13-36)NPY~(18-36)NPY~(3-36)NPY >> (LP)NPY~(LP)(3-36)NPY). The Y5 receptor is also different from the pancreatic polypeptide (PP) receptor (Y4/PP) since [¹²⁵I] PP (rat) does not bind to it.

Although the rank order of affinities is essentially the same when comparing 6B to Y861 and Y555, subtle differences do exist in the IC₅₀ values. It appears that Y861 and Y555 have slightly lower affinities (approximately 2- to 3-fold) for PYY and other PYY analogs, as compared to 6B. In addition, (10-36)NPY and (13-36) have 2- to 4-fold lower affinity for Y861 and Y555.

Nonlinear regression analysis of saturation data for the Y5 receptor yielded a K_d value of 0.27 nM and a receptor density (B_{max}) of about 140 fmol/mg protein in these transiently transfected cells.

Fig. 2 presents the saturation curve for specific binding of [¹²⁵I]PYY to Y5 receptor membranes transiently expressed in COS-7 cells. Membranes were incubated with concentrations of [¹²⁵I]PYY ranging from 0.05 to 5 nM, in the presence or absence of 1 μM PYY. Each point represents the average value of triplicate determinations at each concentration tested. The inset in Fig. 2 shows the corresponding Rosenthal plot of the data.

Table 4

Peptide	EC ₅₀ Values, nM (n Value)	
	293S.Y861.2	293.hY5.sb.8
r/hNPY	6.3 ± 1.9 (3)	0.3 (1)
rPYY	6.5 (2)	ND
r/h(2-36)NPY	21 (2)	ND
r/h(3-36)NPY	ND	6 (1)
r/h(LeuPro)(3-36)NPY	31 ± 39 (3)	23 ± 11 (3)
(32D-Trp)NPY	24 (1)	33 (1)
hPP	1 (1)	5 (1)
rPP	112 (1)	>1000 (1)

Example 5

Isolation of Human Y5 Receptor

10 *Isolation of Human Genomic Clone*

Polymerase chain reaction (PCR) was used to amplify a 375 base pair (bp) coding region of the rat Y5 cDNA clone. The primers for the PCR were:

(+) 5'-TAGGGAACCTGGCCTCCTCC-3' (SEQ ID NO 5) (nucleotides 487-506),

(-) 5'-TCAGAGGGCCATGACTCAAC-3' (SEQ ID NO 6) (nucleotides 843-862).

15 The PCR product was cloned into pCRII vector (Invitrogen) and sequenced. After confirmation by sequencing, the insert was purified from the low melting gel and labeled with digoxigenin-11-dUTP using the random primed method (Boehringer Mannheim, Indianapolis, IN). The labeled probe was used to screen human genomic library.

1x10⁶ independent recombinants were screened from the library. Filter hybridization was carried out in the hybridization buffer containing 6x SSC, 0.1% N-lauroylsarcosine, 0.02% sodium dodecyl sulfate (SDS), 3% blocking reagent (Boehringer Mannheim) and 30% formamide at 37°C overnight. The filters were washed at 37°C in 0.1x SSC, 0.1% SDS and the positive clones were identified by CSPD detection kit according to the manufacturer's protocol (Boehringer Mannheim).

Two positive clones (HG11A and HG19) were isolated from the library. The positive clones were subcloned into pBluescript vector (Stratagene). One clone, h11a, was analyzed by restriction mapping and plasmid Southern blot. Two EcoRV fragments, 2.4 kb and 0.4 kb, were hybridized by the rat Y5 probe. These two DNA fragments were subcloned and sequenced from both ends. DNA sequence analysis was performed using GCG program. The coding region of the human Y5 genomic clone was identified by DNA sequence analysis. This region was amplified by PCR using genomic clone h11A as template and subcloned into pcDNA3 expression vector (Invitrogen) for further studies. The h11A clone has the nucleic acid coding sequence given by SEQ ID NO 5 and the protein that it encodes has the amino acid sequence given by SEQ ID NO 6.

The human Y5 DNA coding region was used to search the sequence similarities in the gene bank. The Y5 coding sequence from nucleotide 821 to the stop codon at position 1338 is nearly identical, *but in an opposite orientation*, to part of the human NPY-Y1 gene (Ball et al, *J. Biol. Chem.* 270, 30102 (1995)). The identical sequence covered the 1C exon promoter, exon 1C, and part of the intron sequences of the NPY-Y1 receptor in an *opposite orientation*. Compared to the published nucleotide sequence, the Y5 coding region has a T insertion at position 1226 and a TG insertion at positions 1235 and 1236.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: Hu Ph.D., Yinghe
McCaleb Ph.D., Michael L.
Bloomquist Ph.D., Brian T.
Flores-Riveros Ph.D., Jaime R.
Cornfield Ph.D., Linda J.

(ii) TITLE OF INVENTION: Neuropeptide Y Receptor and Nucleic Acid
Sequences

(iii) NUMBER OF SEQUENCES: 8

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: McDonnell Boehnen Hulbert & Berghoff
(B) STREET: 300 South Wacker Drive
(C) CITY: Chicago
(D) STATE: IL
(E) COUNTRY: USA
(F) ZIP: 60606

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.30

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: Greenfield Ph.D., Michael S.
(B) REGISTRATION NUMBER: 37,147
(C) REFERENCE/DOCKET NUMBER: 96,149/WH 405

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: (312) 715-1000
(B) TELEFAX: (312) 715-1234

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 2481 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 248..1585

(ix) FEATURE:

(A) NAME/KEY: mat_peptide

(B) LOCATION: 248..1582

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

15	GAATTCTTGG ACTATGGGGG CCGGGAACAG GCGATCTTGA GCCGGGTGTC CGGGGTCTCA	60
	GGGACTGTCA CGTGTTCCTCG AGGTGCTTCT AAAACCCTGG CGGCTCCGGA GCCCTCCTT	120
	CCCACCACCG CCTCCAGGTC CTGCTCCTGC CGCCACCGCT TCCATCTGGA GCAGAAGCGA	180
20	CCGCGCTCAG CCACGTACCC CGGAGTCCAG GCACCCGCAG CGGCCGGGGC ATCCCGAGGA	240
	TTTTAGT ATG GAG TTT AAG CTT GAG GAG CAT TTT AAC AAG ACA TTT GTC	289
	Met Glu Phe Lys Leu Glu Glu His Phe Asn Lys Thr Phe Val	
	1 5 10	
25	ACA GAG AAC AAT ACA GCT GCT GCT CGG AAT GCA GCC TTC CCT GCC TGG	337
	Thr Glu Asn Asn Thr Ala Ala Ala Arg Asn Ala Ala Phe Pro Ala Trp	
	15 20 25 30	
30	GAG GAC TAC AGA GGC AGC GTA GAC GAT TTA CAA TAC TTT CTG ATT GGG	385
	Glu Asp Tyr Arg Gly Ser Val Asp Asp Leu Gln Tyr Phe Leu Ile Gly	
	35 40 45	
35	CTC TAT ACA TTC GTA AGT CTT CTT GGC TTT ATG GGC AAT CTA CCT ATT	433
	Leu Tyr Thr Phe Val Ser Leu Leu Gly Phe Met Gly Asn Leu Pro Ile	
	50 55 60	
40	TTA ATG GCT GTT ATG AAA AAG CGC AAT CAG AAG ACT ACA GTG AAC TTT	481
	Leu Met Ala Val Met Lys Lys Arg Asn Gln Lys Thr Thr Val Asn Phe	
	65 70 75	
45	CTC ATA GGC AAC CTG GCC TTC TCC GAC ATC TTG GTC GTC CTG TTT TGC	529
	Leu Ile Gly Asn Leu Ala Phe Ser Asp Ile Leu Val Val Leu Phe Cys	
	80 85 90	
50	TCC CCT TTC ACC CTG ACC TCT GTC TTG TTG GAT CAG TGG ATG TTT GGC	577
	Ser Pro Phe Thr Leu Thr Ser Val Leu Leu Asp Gln Trp Met Phe Gly	
	95 100 105 110	
55	AAA GCC ATG TGC CAT ATC ATG CCG TTC CTT CAA TGT GTG TCA GTT CTG	625
	Lys Ala Met Cys His Ile Met Pro Phe Leu Gln Cys Val Ser Val Leu	
	115 120 125	
	GTT TCA ACT CTG ATT TTA ATA TCA ATT GCC ATT GTC AGG TAT CAT ATG	673
	Val Ser Thr Leu Ile Leu Ile Ser Ile Ala Ile Val Arg Tyr His Met	
	130 135 140	

	ATA AAG CAC CCT ATT TCT AAC AAT TTA ACG GCA AAC CAT GGC TAC TTC	721
	Ile Lys His Pro Ile Ser Asn Asn Leu Thr Ala Asn His Gly Tyr Phe	
	145 150 155	
5	CTG ATA GCT ACT GTC TGG ACA CTG GGC TTT GCC ATC TGT TCT CCC CTC	769
	Leu Ile Ala Thr Val Trp Thr Leu Gly Phe Ala Ile Cys Ser Pro Leu	
	160 165 170	
10	CCA GTG TTT CAC AGT CTT GTG GAA CTT AAG GAG ACC TTT GGC TCA GCA	817
	Pro Val Phe His Ser Leu Val Glu Leu Lys Glu Thr Phe Gly Ser Ala	
	175 180 185 190	
15	CTG CTG AGT AGC AAA TAT CTC TGT GTT GAG TCA TGG CCC TCT GAT TCA	865
	Leu Leu Ser Ser Lys Tyr Leu Cys Val Glu Ser Trp Pro Ser Asp Ser	
	195 200 205	
20	TAC AGA ATT GCT TTC ACA ATC TCT TTA TTG CTA GTG CAG TAT ATC CTG	913
	Tyr Arg Ile Ala Phe Thr Ile Ser Leu Leu Val Gln Tyr Ile Leu	
	210 215 220	
25	CCT CTA GTA TGT TTA ACG GTA AGT CAT ACC AGC GTC TGC CGA AGC ATA	961
	Pro Leu Val Cys Leu Thr Val Ser His Thr Ser Val Cys Arg Ser Ile	
	225 230 235	
30	AGC TGT GGA TTG TCC CAC AAA GAA AAC AGA CTC GAA GAA AAT GAG ATG	1009
	Ser Cys Gly Leu Ser His Lys Glu Asn Arg Leu Glu Glu Asn Glu Met	
	240 245 250	
35	ATC AAC TTA ACC CTA CAG CCA TCC AAA AAG AGC AGG AAC CAG GCA AAA	1057
	Ile Asn Leu Thr Leu Gln Pro Ser Lys Lys Ser Arg Asn Gln Ala Lys	
	255 260 265 270	
40	ACC CCC AGC ACT CAA AAG TGG AGC TAC TCA TTC ATC AGA AAG CAC AGA	1105
	Thr Pro Ser Thr Gln Lys Trp Ser Tyr Ser Phe Ile Arg Lys His Arg	
	275 280 285	
45	AGG AGG TAC AGC AAG AAG ACG GCC TGT GTC TTA CCC GCC CCA GCA GGA	1153
	Arg Arg Tyr Ser Lys Lys Thr Ala Cys Val Leu Pro Ala Pro Ala Gly	
	290 295 300	
50	CCT TCC CAG GGG AAG CAC CTA GCC GTT CCA GAA AAT CCA GCC TCC GTC	1201
	Pro Ser Gln Gly Lys His Leu Ala Val Pro Glu Asn Pro Ala Ser Val	
	305 310 315	
55	CGT AGC CAG CTG TCG CCA TCC AGT AAG GTC ATT CCA GGG GTC CCA ATC	1249
	Arg Ser Gln Leu Ser Pro Ser Ser Lys Val Ile Pro Gly Val Pro Ile	
	320 325 330	
60	TGC TTT GAG GTG AAA CCT GAA GAA AGC TCA GAT GCT CAT GAG ATG AGA	1297
	Cys Phe Glu Val Lys Pro Glu Glu Ser Ser Asp Ala His Glu Met Arg	
	335 340 345 350	
65	GTC AAG CGT TCC ATC ACT AGA ATA AAA AAG AGA TCT CGA AGT GTT TTC	1345
	Val Lys Arg Ser Ile Thr Arg Ile Lys Lys Arg Ser Arg Ser Val Phe	
	355 360 365	

TAC AGA CTG ACC ATA CTG ATA CTC GTG TTC GCC GTT AGC TGG ATG CCA 1393
 Tyr Arg Leu Thr Ile Leu Ile Leu Val Phe Ala Val Ser Trp Met Pro
 370 375 380

5 CTC CAC GTC TTC CAC GTG GTG ACT GAC TTC AAT GAT AAC TTG ATT TCC 1441
 Leu His Val Phe His Val Val Thr Asp Phe Asn Asp Asn Leu Ile Ser
 385 390 395

AAT AGG CAT TTC AAG CTG GTA TAC TGC ATC TGT CAC TTG TTA GGC ATG 1489
 Asn Arg His Phe Lys Leu Val Tyr Cys Ile Cys His Leu Leu Gly Met
 10 400 405 410

ATG TCC TGT TGT CTA AAT CCG ATC CTA TAT GGT TTC CTT AAT AAT GGT 1537
 Met Ser Cys Cys Leu Asn Pro Ile Leu Tyr Gly Phe Leu Asn Asn Gly
 415 420 425 430

15 ATC AAA GCA GAC TTG AGA GCC CTT ATC CAC TGC CTA CAC ATG TCA TGA 1585
 Ile Lys Ala Asp Leu Arg Ala Leu Ile His Cys Leu His Met Ser *
 435 440 445

20 TTCTCTCTGT GCACCAAAGA GAGAAGAAAC GTGGTAATTG ACACATAATT TATACAGAAG 1645
 TATTCTGGAT CTGAATGCCA GTTCGTAATC TACGTAAGAT CATCTTCATG TTATAATATG 1705
 GTTAATTCAA TCAGTTGTGC AGAGTCAATG TCCATCTAAT ACAATTCAT GTGTTGAAGT 1765

25 AGTTTACATT ATTTTCCATT TTATGTCATT GGTAATAAGT TGAGTGATAC TCTGTGGTTT 1825
 AGTGTAAG AATATAGCTAT CCAAAATTGTT ACGTTGTACA AAAAAATGTAT GAAGTGACAA 1885

30 GTTGTCCCAA AGAGCATTTA ACTACAGATT TAAGGAATTT CTATTATCTG GGTATCTTCA 1945
 TTTCTATTTT ACAGGCTTCT TAACATTTTT TTGTAAAAGT ACAAAAATAT TCAAAAGTCA 2005
 GAACTCTATT ACAGATGTAT GCATAAAAGA TGATTATAAT TTTGTAGGAG AAAGATCTGC 2065

35 TCCTATTAGT GAAGATTGGT AAAATTGTCA GTTTAACCCG GCTGTCCTAC TACTAATATT 2125
 TAATTTTICA AATATGAAAA GGTTTCAGAT TTTGTTTAGA TTTATATCAC ATTAAACACT 2185

40 GTCAAATAAA GGCTGTTTTT ATATGCATCG TTGATGTTCC AAAATGTGAA GTCTAAATGG 2245
 TGTCTGTATT TCCAATTATT AAATAACTTC TAAGATCATT TTTAAAAGTC TGTAGATGGT 2305
 ATGGATAGCT AGTTGTTTGT TAATATAAAG TAAAAGTAGA TAGCTGATTT ATGTTGTACC 2365

45 TATGTCGTAT GTATATTAGG TATCGTGTG TCTCACTAAA GTGAAAGCAA ACGAAAAAAA 2425
 AAAAAAAAAA AAAAAAAAAA AAAAAAAAAA AAAAAAAAAA AAAAAAAAAA AAAAAA 2481

50 (2) INFORMATION FOR SEQ ID NO:2:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 445 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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5  Met Glu Phe Lys Leu Glu Glu His Phe Asn Lys Thr Phe Val Thr Glu
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   Asn Asn Thr Ala Ala Ala Arg Asn Ala Ala Phe Pro Ala Trp Glu Asp
   20             25             30
10  Tyr Arg Gly Ser Val Asp Asp Leu Gln Tyr Phe Leu Ile Gly Leu Tyr
   35             40             45
   Thr Phe Val Ser Leu Leu Gly Phe Met Gly Asn Leu Pro Ile Leu Met
   50             55             60
15  Ala Val Met Lys Lys Arg Asn Gln Lys Thr Thr Val Asn Phe Leu Ile
   65             70             75             80
   Gly Asn Leu Ala Phe Ser Asp Ile Leu Val Val Leu Phe Cys Ser Pro
20             85             90             95
   Phe Thr Leu Thr Ser Val Leu Leu Asp Gln Trp Met Phe Gly Lys Ala
   100            105            110
25  Met Cys His Ile Met Pro Phe Leu Gln Cys Val Ser Val Leu Val Ser
   115            120            125
   Thr Leu Ile Leu Ile Ser Ile Ala Ile Val Arg Tyr His Met Ile Lys
   130            135            140
30  His Pro Ile Ser Asn Asn Leu Thr Ala Asn His Gly Tyr Phe Leu Ile
   145            150            155            160
   Ala Thr Val Trp Thr Leu Gly Phe Ala Ile Cys Ser Pro Leu Pro Val
35             165             170             175
   Phe His Ser Leu Val Glu Leu Lys Glu Thr Phe Gly Ser Ala Leu Leu
   180            185            190
40  Ser Ser Lys Tyr Leu Cys Val Glu Ser Trp Pro Ser Asp Ser Tyr Arg
   195            200            205
   Ile Ala Phe Thr Ile Ser Leu Leu Leu Val Gln Tyr Ile Leu Pro Leu
   210            215            220
45  Val Cys Leu Thr Val Ser His Thr Ser Val Cys Arg Ser Ile Ser Cys
   225            230            235            240
   Gly Leu Ser His Lys Glu Asn Arg Leu Glu Glu Asn Glu Met Ile Asn
50             245             250             255
   Leu Thr Leu Gln Pro Ser Lys Lys Ser Arg Asn Gln Ala Lys Thr Pro
   260            265            270
55  Ser Thr Gln Lys Trp Ser Tyr Ser Phe Ile Arg Lys His Arg Arg Arg
   275            280            285

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Tyr Ser Lys Lys Thr Ala Cys Val Leu Pro Ala Pro Ala Gly Pro Ser
 290 295 300
 5 Gln Gly Lys His Leu Ala Val Pro Glu Asn Pro Ala Ser Val Arg Ser
 305 310 315 320
 Gln Leu Ser Pro Ser Ser Lys Val Ile Pro Gly Val Pro Ile Cys Phe
 325 330 335
 10 Glu Val Lys Pro Glu Glu Ser Ser Asp Ala His Glu Met Arg Val Lys
 340 345 350
 Arg Ser Ile Thr Arg Ile Lys Lys Arg Ser Arg Ser Val Phe Tyr Arg
 355 360 365
 15 Leu Thr Ile Leu Ile Leu Val Phe Ala Val Ser Trp Met Pro Leu His
 370 375 380
 Val Phe His Val Val Thr Asp Phe Asn Asp Asn Leu Ile Ser Asn Arg
 20 385 390 395 400
 His Phe Lys Leu Val Tyr Cys Ile Cys His Leu Leu Gly Met Met Ser
 405 410 415
 25 Cys Cys Leu Asn Pro Ile Leu Tyr Gly Phe Leu Asn Asn Gly Ile Lys
 420 425 430
 Ala Asp Leu Arg Ala Leu Ile His Cys Leu His Met Ser *
 435 440 445

30

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

35

- (A) LENGTH: 2604 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

40

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

45

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 371..1708

50

(ix) FEATURE:

- (A) NAME/KEY: mat_peptide
- (B) LOCATION: 371..1705

55

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GAATTCTTGG ACTATGGGGG CCGGGAACAG GCGATCTTGA GCCGGGTGTC CGGGGTCTCA

60

	GGGACTGTCA CGTGTTCCCG AGGTGCTTCT AAAACCCTGG CGGCTCCGGA GCCCCCTCCTT	120
	CCCACCACCG CCTCCAGGTC CTGCTCCTGC CGCCACCGCT TCCATCTGGA GCAGAAGCGA	180
5	CCGCGCTCAG CCACGTACCC CGGAGTCCAG GCACCCGCAG CGGCCGGGGC ATCCCGAGCT	240
	GGCCATACAC CGGGAGACAG CTGTGCCCTT GGGTTTGCAA GGTGGCTTGG AAGTCAACTG	300
	CCAGTAGGAA ATAGCCATCC ACACACCTGA GTTCCAAGGG GGAAGAAAGA GATTCTTATC	360
10	TGATTTTAGT ATG GAG TTT AAG CTT GAG GAG CAT TTT AAC AAG ACA TTT Met Glu Phe Lys Leu Glu Glu His Phe Asn Lys Thr Phe	409
	1 5 10	
15	GTC ACA GAG AAC AAT ACA GCT GCT GCT CGG AAT GCA GCC TTC CCT GCC Val Thr Glu Asn Asn Thr Ala Ala Ala Arg Asn Ala Ala Phe Pro Ala	457
	15 20 25	
	TGG GAG GAC TAC AGA GGC AGC GTA GAC GAT TTA CAA TAC TTT CTG ATT Trp Glu Asp Tyr Arg Gly Ser Val Asp Asp Leu Gln Tyr Phe Leu Ile	505
20	30 35 40 45	
	GGG CTC TAT ACA TTC GTA AGT CTT CTT GGC TTT ATG GGC AAT CTA CTT Gly Leu Tyr Thr Phe Val Ser Leu Leu Gly Phe Met Gly Asn Leu Leu	553
25	50 55 60	
	ATT TTA ATG GCT GTT ATG AAA AAG CGC AAT CAG AAG ACT ACA GTG AAC Ile Leu Met Ala Val Met Lys Lys Arg Asn Gln Lys Thr Thr Val Asn	601
	65 70 75	
30	TTT CTC ATA GGC AAC CTG GCC TTC TCC GAC ATC TTG GTC GTC CTG TTT Phe Leu Ile Gly Asn Leu Ala Phe Ser Asp Ile Leu Val Val Leu Phe	649
	80 85 90	
35	TGC TCC CCT TTC ACC CTG ACC TCT GTC TTG TTG GAT CAG TGG ATG TTT Cys Ser Pro Phe Thr Leu Thr Ser Val Leu Leu Asp Gln Trp Met Phe	697
	95 100 105	
	GGC AAA GCC ATG TGC CAT ATC ATG CCG TTC CTT CAA TGT GTG TCA GTT Gly Lys Ala Met Cys His Ile Met Pro Phe Leu Gln Cys Val Ser Val	745
40	110 115 120 125	
	CTG GTT TCA ACT CTG ATT TTA ATA TCA ATT GCC ATT GTC AGG TAT CAT Leu Val Ser Thr Leu Ile Leu Ile Ser Ile Ala Ile Val Arg Tyr His	793
45	130 135 140	
	ATG ATA AAG CAC CCT ATT TCT AAC AAT TTA ACG GCA AAC CAT GGC TAC Met Ile Lys His Pro Ile Ser Asn Asn Leu Thr Ala Asn His Gly Tyr	841
	145 150 155	
50	TTC CTG ATA GCT ACT GTC TGG ACA CTG GGC TTT GCC ATC TGT TCT CCC Phe Leu Ile Ala Thr Val Trp Thr Leu Gly Phe Ala Ile Cys Ser Pro	889
	160 165 170	
55	CTC CCA GTG TTT CAC AGT CTT GTG GAA CTT AAG GAG ACC TTT GGC TCA Leu Pro Val Phe His Ser Leu Val Glu Leu Lys Glu Thr Phe Gly Ser	937
	175 180 185	

	GCA CTG CTG AGT AGC AAA TAT CTC TGT GTT GAG TCA TGG CCC TCT GAT	985
	Ala Leu Leu Ser Ser Lys Tyr Leu Cys Val Glu Ser Trp Pro Ser Asp	
	190 195 200 205	
5	TCA TAC AGA ATT GCT TTC ACA ATC TCT TTA TTG CTA GTG CAG TAT ATC	1033
	Ser Tyr Arg Ile Ala Phe Thr Ile Ser Leu Leu Val Gln Tyr Ile	
	210 215 220	
10	CTG CCT CTA GTA TGT TTA ACG GTA AGT CAT ACC AGC GTC TGC CGA AGC	1081
	Leu Pro Leu Val Cys Leu Thr Val Ser His Thr Ser Val Cys Arg Ser	
	225 230 235	
	ATA AGC TGT GGA TTG TCC CAC AAA GAA AAC AGA CTC GAA GAA AAT GAG	1129
	Ile Ser Cys Gly Leu Ser His Lys Glu Asn Arg Leu Glu Glu Asn Glu	
	240 245 250	
15	ATG ATC AAC TTA ACC CTA CAG CCA TCC AAA AAG AGC AGG AAC CAG GCA	1177
	Met Ile Asn Leu Thr Leu Gln Pro Ser Lys Lys Ser Arg Asn Gln Ala	
	255 260 265	
20	AAA ACC CCC AGC ACT CAA AAG TGG AGC TAC TCA TTC ATC AGA AAG CAC	1225
	Lys Thr Pro Ser Thr Gln Lys Trp Ser Tyr Ser Phe Ile Arg Lys His	
	270 275 280 285	
25	AGA AGG AGG TAC AGC AAG AAG ACG GCC TGT GTC TTA CCC GCC CCA GCA	1273
	Arg Arg Arg Tyr Ser Lys Lys Thr Ala Cys Val Leu Pro Ala Pro Ala	
	290 295 300	
30	GGA CCT TCC CAG GGG AAG CAC CTA GCC GTT CCA GAA AAT CCA GCC TCC	1321
	Gly Pro Ser Gln Gly Lys His Leu Ala Val Pro Glu Asn Pro Ala Ser	
	305 310 315	
35	GTC CGT AGC CAG CTG TCG CCA TCC AGT AAG GTC ATT CCA GGG GTC CCA	1369
	Val Arg Ser Gln Leu Ser Pro Ser Ser Lys Val Ile Pro Gly Val Pro	
	320 325 330	
40	ATC TGC TTT GAG GTG AAA CCT GAA GAA AGC TCA GAT GCT CAT GAG ATG	1417
	Ile Cys Phe Glu Val Lys Pro Glu Glu Ser Ser Asp Ala His Glu Met	
	335 340 345	
	AGA GTC AAG CGT TCC ATC ACT AGA ATA AAA AAG AGA TCT CGA AGT GTT	1465
	Arg Val Lys Arg Ser Ile Thr Arg Ile Lys Lys Arg Ser Arg Ser Val	
	350 355 360 365	
45	TTC TAC AGA CTG ACC ATA CTG ATA CTC GTG TTC GCC GTT AGC TGG ATG	1513
	Phe Tyr Arg Leu Thr Ile Leu Ile Leu Val Phe Ala Val Ser Trp Met	
	370 375 380	
50	CCA CTC CAC GTC TTC CAC GTG GTG ACT GAC TTC AAT GAT AAC TTG ATT	1561
	Pro Leu His Val Phe His Val Val Thr Asp Phe Asn Asp Asn Leu Ile	
	385 390 395	
55	TCC AAT AGG CAT TTC AAG CTG GTA TAC TGC ATC TGT CAC TTG TTA GGC	1609
	Ser Asn Arg His Phe Lys Leu Val Tyr Cys Ile Cys His Leu Leu Gly	
	400 405 410	

ATG ATG TCC TGT TGT CTA AAT CCG ATC CTA TAT GGT TTC CTT AAT AAT 1657
 Met Met Ser Cys Cys Leu Asn Pro Ile Leu Tyr Gly Phe Leu Asn Asn
 415 420 425
 5 GGT ATC AAA GCA GAC TTG AGA GCC CTT ATC CAC TGC CTA CAC ATG TCA 1705
 Gly Ile Lys Ala Asp Leu Arg Ala Leu Ile His Cys Leu His Met Ser
 430 435 440 445
 10 TGA TTCTCTCTGT GCACCAAAGA GAGAAGAAAC GTGGTAATTG ACACATAATT 1758
 *
 TATACAGAAG TATTCTGGAT CTGAATGCCA GTTCGTAATC TACGTAAGAT CATCTTCATG 1818
 15 TTATAATATG GTTAATTCAA TCAGTTGTGC AGAGTCAATG TCCATCTAAT ACAATTTTCAT 1878
 GTGTTGAAGT AGTTTACATT ATTTTCCATT TTATGTCATT GGTAATAAGT TGAGTGATAC 1938
 TCTGTGGTTT AGTGTA AAAAG ATATAGCTAT CCAAATTGTT ACGTTGTACA AAAAATGTAT 1998
 20 GAAGTGACAA GTTGTCCCAA AGAGCATTTA ACTACAGATT TAAGGAATTT CTATTATCTG 2058
 GGTATCTTCA TTTCTATTTC ACAGGCTTCT TAACATTTTT TTGTAAAAGT ACAAAAATAT 2118
 25 TCAAAAGTCA GAACTCTATT ACAGATGTAT GCATAAAAGA TGATTATAAT TTTGTAGGAG 2178
 AAAGATCTGC TCCTATTAGT GAAGATTGGT AAAATTGTCA GTTTAACCCG GCTGTCCTAC 2238
 TACTAATATT TAATTTTTCA AATATGAAAA GGTTTCAGAT TTTGTTTAGA TTTATATCAC 2298
 30 ATTAACACT GTCAAATAAA GGCTGTTTTT ATATGCATCG TTGATGTTCC AAAATGTGAA 2358
 GTCTAAATGG TGTCTGTATT TCCAATTATT AAATAACTTC TAAGATCATT TTTAAAAGTC 2418
 35 TGTAGATGGT ATGGATAGCT AGTTGTTTGT TAATATAAAG TAAAAGTAGA TAGCTGATTT 2478
 ATGTTGTACC TATGTCGTAT GTATATTAGG TATCGTGTTG TCTCACTAAA GTGAAAGCAA 2538
 ACGAAAAAAA AAAAAAAAAA AAAAAAAAAA AAAAAAAAAA AAAAAAAAAA AAAAAAAAAA 2598
 40 AAAAAA 2604

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 445 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

55 Met Glu Phe Lys Leu Glu Glu His Phe Asn Lys Thr Phe Val Thr Glu
 1 5 10 15

Asn Asn Thr Ala Ala Ala Arg Asn Ala Ala Phe Pro Ala Trp Glu Asp
 20 25 30
 Tyr Arg Gly Ser Val Asp Asp Leu Gln Tyr Phe Leu Ile Gly Leu Tyr
 5 35 40 45
 Thr Phe Val Ser Leu Leu Gly Phe Met Gly Asn Leu Leu Ile Leu Met
 50 55 60
 10 Ala Val Met Lys Lys Arg Asn Gln Lys Thr Thr Val Asn Phe Leu Ile
 65 70 75 80
 Gly Asn Leu Ala Phe Ser Asp Ile Leu Val Val Leu Phe Cys Ser Pro
 85 90 95
 15 Phe Thr Leu Thr Ser Val Leu Leu Asp Gln Trp Met Phe Gly Lys Ala
 100 105 110
 Met Cys His Ile Met Pro Phe Leu Gln Cys Val Ser Val Leu Val Ser
 20 115 120 125
 Thr Leu Ile Leu Ile Ser Ile Ala Ile Val Arg Tyr His Met Ile Lys
 130 135 140
 25 His Pro Ile Ser Asn Asn Leu Thr Ala Asn His Gly Tyr Phe Leu Ile
 145 150 155 160
 Ala Thr Val Trp Thr Leu Gly Phe Ala Ile Cys Ser Pro Leu Pro Val
 165 170 175
 30 Phe His Ser Leu Val Glu Leu Lys Glu Thr Phe Gly Ser Ala Leu Leu
 180 185 190
 35 Ser Ser Lys Tyr Leu Cys Val Glu Ser Trp Pro Ser Asp Ser Tyr Arg
 195 200 205
 Ile Ala Phe Thr Ile Ser Leu Leu Leu Val Gln Tyr Ile Leu Pro Leu
 210 215 220
 40 Val Cys Leu Thr Val Ser His Thr Ser Val Cys Arg Ser Ile Ser Cys
 225 230 235 240
 Gly Leu Ser His Lys Glu Asn Arg Leu Glu Glu Asn Glu Met Ile Asn
 45 245 250 255
 Leu Thr Leu Gln Pro Ser Lys Lys Ser Arg Asn Gln Ala Lys Thr Pro
 260 265 270
 50 Ser Thr Gln Lys Trp Ser Tyr Ser Phe Ile Arg Lys His Arg Arg Arg
 275 280 285
 Tyr Ser Lys Lys Thr Ala Cys Val Leu Pro Ala Pro Ala Gly Pro Ser
 290 295 300
 55 Gln Gly Lys His Leu Ala Val Pro Glu Asn Pro Ala Ser Val Arg Ser
 305 310 315 320

Gln Leu Ser Pro Ser Ser Lys Val Ile Pro Gly Val Pro Ile Cys Phe
 325 330 335

5 Glu Val Lys Pro Glu Glu Ser Ser Asp Ala His Glu Met Arg Val Lys
 340 345 350

Arg Ser Ile Thr Arg Ile Lys Lys Arg Ser Arg Ser Val Phe Tyr Arg
 355 360 365

10 Leu Thr Ile Leu Ile Leu Val Phe Ala Val Ser Trp Met Pro Leu His
 370 375 380

Val Phe His Val Val Thr Asp Phe Asn Asp Asn Leu Ile Ser Asn Arg
 385 390 395 400

15 His Phe Lys Leu Val Tyr Cys Ile Cys His Leu Leu Gly Met Met Ser
 405 410 415

20 Cys Cys Leu Asn Pro Ile Leu Tyr Gly Phe Leu Asn Asn Gly Ile Lys
 420 425 430

Ala Asp Leu Arg Ala Leu Ile His Cys Leu His Met Ser *

435 440 445

25 (2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- 30 (A) LENGTH: 1338 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: genomic DNA

35 (iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

- 40 (A) NAME/KEY: CDS
 (B) LOCATION: 1..1338

(ix) FEATURE:

- 45 (A) NAME/KEY: mat_peptide
 (B) LOCATION: 1..1335

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

50 ATG GAT TTA GAG CTC GAC GAG TAT TAT AAC AAG ACA CTT GCC ACA GAG 48
 Met Asp Leu Glu Leu Asp Glu Tyr Tyr Asn Lys Thr Leu Ala Thr Glu
 1 5 10 15

AAT AAT ACT GCT GCC ACT CGG AAT TCT GAT TTC CCA GTC TGG GAT GAC 96
 55 Asn Asn Thr Ala Ala Thr Arg Asn Ser Asp Phe Pro Val Trp Asp Asp
 20 25 30

	TAT AAA AGC AGT GTA GAT GAC TTA CAG TAT TTT CTG ATT GGG CTC TAT	144
	Tyr Lys Ser Ser Val Asp Asp Leu Gln Tyr Phe Leu Ile Gly Leu Tyr	
	35 40 45	
5	ACA TTT GTA AGT CTT CTT GGC TTT ATG GGG AAT CTA CTT ATT TTA ATG	192
	Thr Phe Val Ser Leu Leu Gly Phe Met Gly Asn Leu Leu Ile Leu Met	
	50 55 60	
10	GCT CTC ATG AAA AAG CGT AAT CAG AAG ACT ACG GTA AAC TTC CTC ATA	240
	Ala Leu Met Lys Lys Arg Asn Gln Lys Thr Thr Val Asn Phe Leu Ile	
	65 70 75 80	
15	GGC AAT CTG GCC TTT TCT GAT ATC TTG GTT GTG CTG TTT TGC TCA CCT	288
	Gly Asn Leu Ala Phe Ser Asp Ile Leu Val Val Leu Phe Cys Ser Pro	
	85 90 95	
20	TTC ACA CTG ACG TCT GTC TTG CTG GAT CAG TGG ATG TTT GGC AAA GTC	336
	Phe Thr Leu Thr Ser Val Leu Leu Asp Gln Trp Met Phe Gly Lys Val	
	100 105 110	
25	ATG TGC CAT ATT ATG CCT TTT CTT CAA TGT GTG TCA GTT TTG GTT TCA	384
	Met Cys His Ile Met Pro Phe Leu Gln Cys Val Ser Val Leu Val Ser	
	115 120 125	
30	ACT TTA ATT TTA ATA TCA ATT GCC ATT GTC AGG TAT CAT ATG ATA AAA	432
	Thr Leu Ile Leu Ile Ser Ile Ala Ile Val Arg Tyr His Met Ile Lys	
	130 135 140	
35	CAT CCC ATA TCT AAT AAT TTA ACA GCA AAC CAT GGC TAC TTT CTG ATA	480
	His Pro Ile Ser Asn Asn Leu Thr Ala Asn His Gly Tyr Phe Leu Ile	
	145 150 155 160	
40	GCT ACT GTC TGG ACA CTA GGT TTT GCC ATC TGT TCT CCC CTT CCA GTG	528
	Ala Thr Val Trp Thr Leu Gly Phe Ala Ile Cys Ser Pro Leu Pro Val	
	165 170 175	
45	TTT CAC AGT CTT GTG GAA CTT CAA GAA ACA TTT GGT TCA GCA TTG CTG	576
	Phe His Ser Leu Val Glu Leu Gln Glu Thr Phe Gly Ser Ala Leu Leu	
	180 185 190	
50	AGC AGC AGG TAT TTA TGT GTT GAG TCA TGG CCA TCT GAT TCA TAC AGA	624
	Ser Ser Arg Tyr Leu Cys Val Glu Ser Trp Pro Ser Asp Ser Tyr Arg	
	195 200 205	
55	ATT GCC TTT ACT ATC TCT TTA TTG CTA GTT CAG TAT ATT CTG CCC TTA	672
	Ile Ala Phe Thr Ile Ser Leu Leu Leu Val Gln Tyr Ile Leu Pro Leu	
	210 215 220	
60	GTT TGT CTT ACT GTA AGT CAT ACA AGT GTC TGC AGA AGT ATA AGC TGT	720
	Val Cys Leu Thr Val Ser His Thr Ser Val Cys Arg Ser Ile Ser Cys	
	225 230 235 240	
65	GGA TTG TCC AAC AAA GAA AAC AGA CTT GAA GAA AAT GAG ATG ATC AAC	768
	Gly Leu Ser Asn Lys Glu Asn Arg Leu Glu Glu Asn Glu Met Ile Asn	
	245 250 255	

	TTA ACT CTT CAT CCA TCC AAA AAG AGT GGG CCT CAG GTG AAA CTC TCT	816
	Leu Thr Leu His Pro Ser Lys Lys Ser Gly Pro Gln Val Lys Leu Ser	
	260 265 270	
5	GGC AGC CAT AAA TGG AGT TAT TCA TTC ATC AAA AAA CAC AGA AGA AGA	864
	Gly Ser His Lys Trp Ser Tyr Ser Phe Ile Lys Lys His Arg Arg Arg	
	275 280 285	
10	TAT AGC AAG AAG ACA GCA TGT GTG TTA CCT GCT CCA GAA AGA CCT TCT	912
	Tyr Ser Lys Lys Thr Ala Cys Val Leu Pro Ala Pro Glu Arg Pro Ser	
	290 295 300	
15	CAA GAG AAC CAC TCC AGA ATA CTT CCA GAA AAC TTT GGC TCT GTA AGA	960
	Gln Glu Asn His Ser Arg Ile Leu Pro Glu Asn Phe Gly Ser Val Arg	
	305 310 315 320	
	AGT CAG CTC TCT TCA TCC AGT AAG TTC ATA CCA GGG GTC CCC ACT TGC	1008
	Ser Gln Leu Ser Ser Ser Ser Lys Phe Ile Pro Gly Val Pro Thr Cys	
	325 330 335	
20	TTT GAG ATA AAA CCT GAA GAA AAT TCA GAT GTT CAT GAA TTG AGA GTA	1056
	Phe Glu Ile Lys Pro Glu Glu Asn Ser Asp Val His Glu Leu Arg Val	
	340 345 350	
25	AAA CGT TCT GTT ACA AGA ATA AAA AAG AGA TCT CGA AGT GTT TTC TAC	1104
	Lys Arg Ser Val Thr Arg Ile Lys Lys Arg Ser Arg Ser Val Phe Tyr	
	355 360 365	
30	AGA CTG ACC ATA CTG ATA TTA GTA TTT GCT GTT AGT TGG ATG CCA CTA	1152
	Arg Leu Thr Ile Leu Ile Leu Val Phe Ala Val Ser Trp Met Pro Leu	
	370 375 380	
35	CAC CTT TTC CAT GTG GTA ACT GAT TTT AAT GAC AAT CTT ATT TCA AAT	1200
	His Leu Phe His Val Val Thr Asp Phe Asn Asp Asn Leu Ile Ser Asn	
	385 390 395 400	
	AGG CAT TTC AAG TTG GTG TAT TGC ATT TGT CAT TTG TTG GGC ATG ATG	1248
	Arg His Phe Lys Leu Val Tyr Cys Ile Cys His Leu Leu Gly Met Met	
	405 410 415	
40	TCC TGT TGT CTT AAT CCA ATT CTA TAT GGG TTT CTT AAT AAT GGG ATT	1296
	Ser Cys Cys Leu Asn Pro Ile Leu Tyr Gly Phe Leu Asn Asn Gly Ile	
	420 425 430	
45	AAA GCT GAT TTA GTG TCC CTT ATA CAC TGT CTT CAT ATG TAA	1338
	Lys Ala Asp Leu Val Ser Leu Ile His Cys Leu His Met *	
	435 440 445	

50 (2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 445 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Asp Leu Glu Leu Asp Glu Tyr Tyr Asn Lys Thr Leu Ala Thr Glu
 1 5 10 15
 5 Asn Asn Thr Ala Ala Thr Arg Asn Ser Asp Phe Pro Val Trp Asp Asp
 20 25 30
 10 Tyr Lys Ser Ser Val Asp Asp Leu Gln Tyr Phe Leu Ile Gly Leu Tyr
 35 40 45
 Thr Phe Val Ser Leu Leu Gly Phe Met Gly Asn Leu Leu Ile Leu Met
 50 55 60
 15 Ala Leu Met Lys Lys Arg Asn Gln Lys Thr Thr Val Asn Phe Leu Ile
 65 70 75 80
 Gly Asn Leu Ala Phe Ser Asp Ile Leu Val Val Leu Phe Cys Ser Pro
 85 90 95
 20 Phe Thr Leu Thr Ser Val Leu Leu Asp Gln Trp Met Phe Gly Lys Val
 100 105 110
 25 Met Cys His Ile Met Pro Phe Leu Gln Cys Val Ser Val Leu Val Ser
 115 120 125
 Thr Leu Ile Leu Ile Ser Ile Ala Ile Val Arg Tyr His Met Ile Lys
 130 135 140
 30 His Pro Ile Ser Asn Asn Leu Thr Ala Asn His Gly Tyr Phe Leu Ile
 145 150 155 160
 Ala Thr Val Trp Thr Leu Gly Phe Ala Ile Cys Ser Pro Leu Pro Val
 165 170 175
 35 Phe His Ser Leu Val Glu Leu Gln Glu Thr Phe Gly Ser Ala Leu Leu
 180 185 190
 40 Ser Ser Arg Tyr Leu Cys Val Glu Ser Trp Pro Ser Asp Ser Tyr Arg
 195 200 205
 Ile Ala Phe Thr Ile Ser Leu Leu Leu Val Gln Tyr Ile Leu Pro Leu
 210 215 220
 45 Val Cys Leu Thr Val Ser His Thr Ser Val Cys Arg Ser Ile Ser Cys
 225 230 235 240
 Gly Leu Ser Asn Lys Glu Asn Arg Leu Glu Glu Asn Glu Met Ile Asn
 245 250 255
 50 Leu Thr Leu His Pro Ser Lys Lys Ser Gly Pro Gln Val Lys Leu Ser
 260 265 270
 55 Gly Ser His Lys Trp Ser Tyr Ser Phe Ile Lys Lys His Arg Arg Arg
 275 280 285

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Asp Leu Glu Leu Asp Glu Tyr Tyr Asn Lys Thr Leu Ala Thr Glu
 1 5 10 15
 5 Asn Asn Thr Ala Ala Thr Arg Asn Ser Asp Phe Pro Val Trp Asp Asp
 20 25 30
 10 Tyr Lys Ser Ser Val Asp Asp Leu Gln Tyr Phe Leu Ile Gly Leu Tyr
 35 40 45
 Thr Phe Val Ser Leu Leu Gly Phe Met Gly Asn Leu Leu Ile Leu Met
 50 55 60
 15 Ala Leu Met Lys Lys Arg Asn Gln Lys Thr Thr Val Asn Phe Leu Ile
 65 70 75 80
 Gly Asn Leu Ala Phe Ser Asp Ile Leu Val Val Leu Phe Cys Ser Pro
 85 90 95
 20 Phe Thr Leu Thr Ser Val Leu Leu Asp Gln Trp Met Phe Gly Lys Val
 100 105 110
 Met Cys His Ile Met Pro Phe Leu Gln Cys Val Ser Val Leu Val Ser
 115 120 125
 25 Thr Leu Ile Leu Ile Ser Ile Ala Ile Val Arg Tyr His Met Ile Lys
 130 135 140
 30 His Pro Ile Ser Asn Asn Leu Thr Ala Asn His Gly Tyr Phe Leu Ile
 145 150 155 160
 Ala Thr Val Trp Thr Leu Gly Phe Ala Ile Cys Ser Pro Leu Pro Val
 165 170 175
 35 Phe His Ser Leu Val Glu Leu Gln Glu Thr Phe Gly Ser Ala Leu Leu
 180 185 190
 Ser Ser Arg Tyr Leu Cys Val Glu Ser Trp Pro Ser Asp Ser Tyr Arg
 195 200 205
 40 Ile Ala Phe Thr Ile Ser Leu Leu Val Gln Tyr Ile Leu Pro Leu
 210 215 220
 45 Val Cys Leu Thr Val Ser His Thr Ser Val Cys Arg Ser Ile Ser Cys
 225 230 235 240
 Gly Leu Ser Asn Lys Glu Asn Arg Leu Glu Glu Asn Glu Met Ile Asn
 245 250 255
 50 Leu Thr Leu His Pro Ser Lys Lys Ser Gly Pro Gln Val Lys Leu Ser
 260 265 270
 Gly Ser His Lys Trp Ser Tyr Ser Phe Ile Lys Lys His Arg Arg Arg
 275 280 285
 55

Tyr Ser Lys Lys Thr Ala Cys Val Leu Pro Ala Pro Glu Arg Pro Ser
 290 295 300
 5 Gln Glu Asn His Ser Arg Ile Leu Pro Glu Asn Phe Gly Ser Val Arg
 305 310 315 320
 Ser Gln Leu Ser Ser Ser Ser Lys Phe Ile Pro Gly Val Pro Thr Cys
 325 330 335
 10 Phe Glu Ile Lys Pro Glu Glu Asn Ser Asp Val His Glu Leu Arg Val
 340 345 350
 Lys Arg Ser Val Thr Arg Ile Lys Lys Arg Ser Arg Ser Val Phe Tyr
 355 360 365
 15 Arg Leu Thr Ile Leu Ile Leu Val Phe Ala Val Ser Trp Met Pro Leu
 370 375 380
 His Leu Phe His Val Val Thr Asp Phe Asn Asp Asn Leu Ile Ser Asn
 20 385 390 395 400
 Arg His Phe Lys Leu Val Tyr Cys Ile Cys His Leu Leu Gly Met Met
 405 410 415
 25 Ser Cys Cys Leu Asn Pro Ile Leu Tyr Gly Phe Leu Asn Asn Gly Ile
 420 425 430
 Lys Ala Asp Leu Val Ser Leu Ile His Cys Leu His Met *
 435 440 445

30 (2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 35 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

40

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

45 TAGGGAACCT GGCCTCCTCC

20

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

50 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

55 (ii) MOLECULE TYPE: other nucleic acid

WO 97/37998

PCT/US97/05781

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:8:

TCAGAGGGCC ATGACTCAAC

20

We claim:

1. An isolated nucleic acid encoding a neuropeptide Y receptor comprising a molecule having a nucleotide sequence substantially the same as SEQ ID NO 1, SEQ ID NO 3, or SEQ ID NO 5.
2. A homogenous composition of the neuropeptide Y receptor comprising a molecule having an amino acid sequence substantially the same as SEQ ID NO 2, SEQ ID NO 4, or SEQ ID NO 6.
3. A vector comprising the nucleic acid according to claim 1.
4. A vector according to claim 3 adapted for expression in a cell further comprising regulatory elements necessary for expression of the nucleic acid operatively linked to the nucleic acid to enable expression of the nucleic acid.
5. A vector according to claim 4 wherein the cell is a mammalian cell.
6. A vector according to claim 5 wherein the cell is a human 293 cell.
7. A vector according to claim 4 that is a plasmid.
8. A vector according to claim 7 wherein the plasmid is the pBluescript plasmid.
9. A vector according to claim 7 wherein the plasmid is the pcDNA3 plasmid.
10. A vector according to claim 3 which is self-replicating.
11. A cell transformed with the nucleic acid according to claim 1 that expresses the nucleic acid.
12. A cell according to claim 11 that is a bacterium cell, an insect cell, or a yeast cell.
13. A cell according to claim 11 that is a mammalian cell.
14. A cell according to claim 13 that is a human 293 cell.

15. A nucleic acid probe comprising a nucleic acid complementary to the nucleic acid according to claim 1.
16. An antisense oligonucleotide having a sequence complementary to the nucleic acid according to claim 1 and that inhibits expression of the nucleic acid.
17. A membrane or membrane preparation comprising a membrane or portion thereof of a cell expressing a nucleic acid having a nucleotide sequence substantially the same as SEQ ID NO 1, SEQ ID NO 3, or SEQ ID NO 5.
18. A membrane or membrane preparation according to claim 17 wherein the cell is a mammalian cell.
19. An antibody or fragment thereof that is immunologically reactive to a mammalian Y5 receptor.
20. An antibody or fragment according to claim 19 thereof wherein the mammalian Y5 receptor has an amino acid sequence substantially the same as one chosen from the group consisting of SEQ ID NO 2, SEQ ID NO 4, and SEQ ID NO 6.
21. An antibody or fragment thereof according to claim 19 that is a monoclonal antibody.
22. An antibody or fragment thereof according to claim 20 that is a monoclonal antibody.
23. A cell line producing an antibody according to claim 19.
24. A cell line producing an antibody according to claim 20.
25. A cell line producing an antibody according to claim 21.
26. A cell line producing an antibody according to claim 22.
27. An epitope of a mammalian Y5 receptor wherein the epitope is immunologically reactive to the antibody or fragment thereof according to claim 19.
28. An epitope of a mammalian Y5 receptor wherein the epitope is immunologically reactive to the antibody or fragment thereof according to claim 20.

29. An epitope of a mammalian Y5 receptor wherein the epitope is immunologically reactive to the antibody or fragment thereof according to claim 21.
30. An epitope of a mammalian Y5 receptor wherein the epitope is immunologically reactive to the antibody or fragment thereof according to claim 22.
31. A method of producing a neuropeptide Y receptor comprising culturing a cell according to claim 11 and recovering the receptor expressed by the cell.
32. A method of producing a neuropeptide Y receptor comprising culturing a cell according to claim 12 and recovering the receptor expressed by the cell.
33. A method of producing a neuropeptide Y receptor comprising culturing a cell according to claim 13 and recovering the receptor expressed by the cell.
34. A method of producing a neuropeptide Y receptor comprising culturing a cell according to claim 14 and recovering the receptor expressed by the cell.
35. A method of identifying a neuropeptide Y agonist or antagonist comprising contacting a potential agonist or antagonist molecule with a membrane or membrane preparation according to claim 17.
36. A method of identifying a neuropeptide Y agonist or antagonist comprising contacting a potential agonist or antagonist molecule with a membrane or membrane preparation according to claim 18.
37. A neuropeptide Y antagonist comprising a compound identified according to claim 36.
38. A method of suppressing the appetite of a mammal comprising administering to the mammal an appetite suppressing amount of a neuropeptide Y antagonist according to claim 37.
39. A method of suppressing the appetite of a mammal according to claim 38 wherein the amount of antagonist is from about 0.01 to about 100 mg/kg.
40. A pharmaceutical composition comprising an effective appetite suppressing amount of an antagonist according to claim 37 together with a pharmaceutically acceptable carrier.

41. A neuropeptide Y agonist comprising a compound identified according to claim 36.
42. A method of stimulating the appetite of a mammal comprising administering to the mammal an appetite stimulating amount of a neuropeptide Y agonist according to claim 41.
43. A method of stimulating the appetite of a mammal according to claim 42 wherein the amount of agonist is from about 0.01 to about 100 mg/kg.
44. A pharmaceutical composition comprising an effective appetite stimulating amount of an agonist according to claim 41 together with a pharmaceutically acceptable carrier.
45. A non-human transgenic mammal that expresses the nucleic acid having a sequence substantially the same one chosen from the group consisting of SEQ ID NO 1, SEQ ID NO 3, and SEQ ID NO 5.

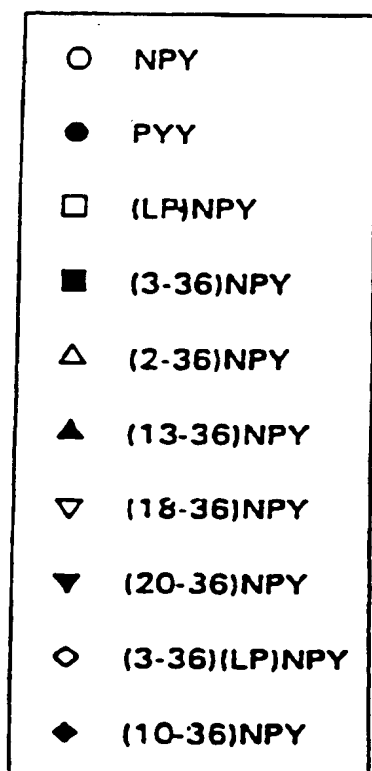
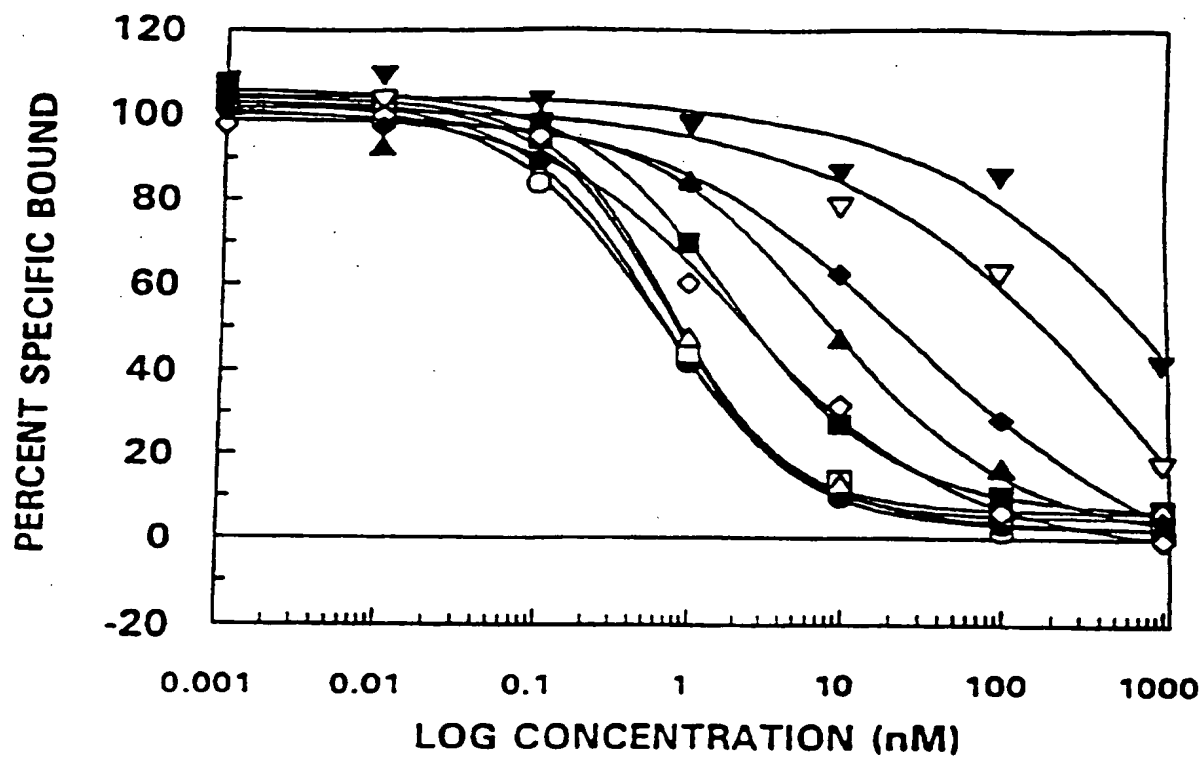
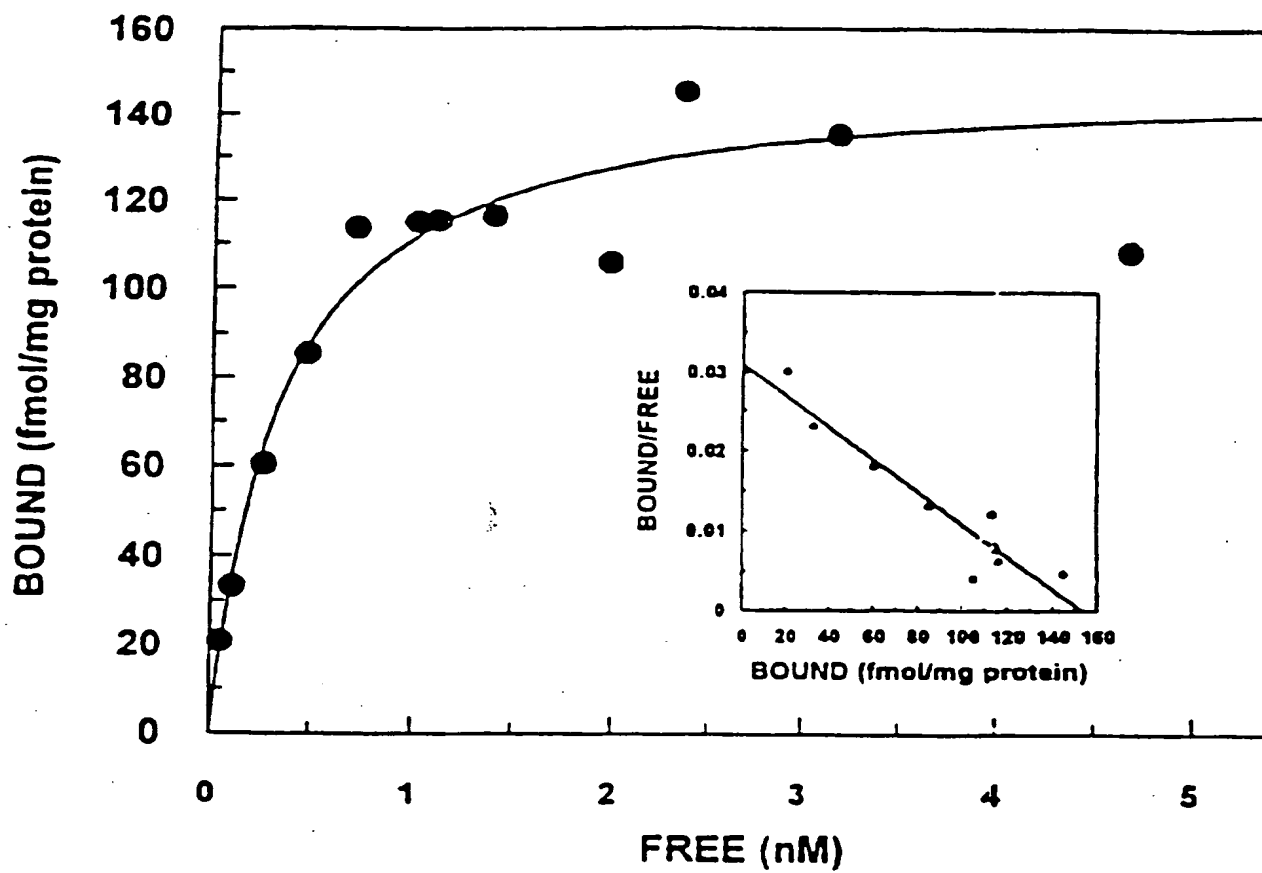


Fig. 1

**Fig. 2**